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Effect of interface-active proteins on the salt crystal size in waterborne hybrid materials

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Abstract

Aqueous processes yielding hybrid or composite materials are widespread in natural environments and their control is fundamental for a multiplicity of living organisms. Their design and in vitro engineering require knowledge about the spatiotemporal evolution of the interactions between the involved liquid and solid phases and, especially, the interphases governing the development of adhesion during solidification. The present study illustrates the effects of distinct proteins on the precipitation of sodium chloride encompassing the size, shape and distribution of halite crystals formed during the drying of droplets containing equally concentrated saline protein solutions. The precipitates obtained from aqueous sodium chloride formulations buffered with tris(hydroxymethyl)aminomethane (Tris) contained either bovine serum albumin (BSA), fibrinogen or collagen and were characterized with respect to their structure and composition using optical and electron microscopy as well as x-ray analysis. The acquired findings highlight that depending on the protein type present during droplet drying the halite deposits predominantly exhibit cubic or polycrystalline dendritic structures. Based on the phenomenological findings, it is suggested that the formation of the interphase between the growing salt phase and the highly viscous saline aqueous jelly phase containing protein governs not only the material transport in the liquid but also the material exchange between the solid and liquid phases.

Keywords: Protein-based composites, Sodium chloride, Bovine serum albumin, Collagen, Fibrinogen, Crystallization, Spatiotemporal development of adhesion, Damköhler number

Introduction

Natural hybrid or composite materials that are composed in aqueous environments and consist of organic and inorganic components or constituents are highly visible and frequent. Seashells, nacre, and bone are protein-based composite materials known for their high functionality at low weight, and their remains from geological eras long ago still shape the landscape in many regions of the world [1]. Moreover, understanding biomineralization processes and characterizing the properties of biomineralized organic/inorganic hybrid materials provide inspiration for materials development following biomimetic principles [2].



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Protein-based composite materials with multi-level hierarchical structures and manifold functions form the basis of many living organisms [3]. Studies of prokaryotes, probably the oldest life-form on Earth, reveal that biomineralization is found in halophiles living in environments governed by high pH and high salt concentrations [4]. These extremophiles are able to attach ions to their outer surface to form minerals [5, 6]. This ability is attributed to a two-dimensional protein assembly, the so-called S-layer (surface layer) of such bacteria and archaea [5]. While the vast majority of biominerals found on Earth are inorganic compounds with oxoanions like carbonates, silicates or phosphates, the crystallization of the evaporite mineral halite, which is based on sodium chloride, is reportedly influenced by the presence of organic entities. These entities include, synthetically produced nanoparticles proposed as an analog to protocellular material [7], surface layers of Haloarcula strain SP8807 or the presence of halobacteria [8], the synthetic peptide ATEA [9], and also gelatin [10, 11]. The crystallization of calcium oxalate or phosphate salts during urolithogenesis in urine or in vitro may be influenced by both dissolved salts and the presence of proteins that contribute to the organic matrix of urinary stones up to several millimeters in width [12, 13]. Hence, in addition to illustrating the effect of organic compounds on salt crystallization, the reciprocal effect of salts on the solidification of organic materials can be highlighted. For example, many proteins precipitate in the presence of highly concentrated salt solutions, like sodium chloride. This effect is known as salting out and its distinctness is reflected by the position of the salt-forming ions in the Hofmeister series [14], which elucidate the tendency of proteins to form aggregates in the presence of different ions [15]. Sodium chloride has also been reported to govern the solubility of lysozyme, α -chymotrypsin and BSA [16]; drive the nucleation and the polymorphic transformation of the amino acid glycine [17]; influence specific attractive interactions relevant for the crystallization and precipitation behaviour of proteins like lysozyme [18]; and affect the aggregation of pork myofibrillar protein [19]. Moreover, sodium chloride can influence the tendency of proteins to hierarchically assemble into superstructures like fibers, such as with spider silk protein [20], the extracellular matrix protein collagen [21], and the blood plasma protein fibrinogen [22]. Submicroscopic protein crystals based on dense aggregates of macromolecular protein units were found in equine chondrocyte cells [23], and, nanoparticulate or crystalline structures composed of proteins and carbohydrates, and containing traces of metals like Na, K, Mg, Ca, Al, and Si were identified in secretions [24].

Notably, the development of adhesion during natural processes occurring at dynamic interphases around solids is in the focus of a growing research area. Acquiring knowledge relating to anti-freeze proteins (AFP) [25, 26] or peptide-based bioadhesives [27] in terms of composition, structural design, and interaction with surfaces is essential for the implementation of underlying concepts based on biochemical and mechanical principles in trendsetting medical and technical applications [28, 29]. In the view of adhesion research, solid formation from an originally continuous fluid phase is an essential scenario during physical hardening [30] and even chemical curing [31]. Rätzke et al. highlighted that a causal sequence of process steps following nucleation and growth may start from sites governing the formation of interphases between the fluid and hardened phases [31]. These locally developing interphases may be perceived as the centerpiece of growth around centers. In these boundary regions, a balance evolves between

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the arrangement of material entities upon solid formation and the transport of entities towards the shifting solid/fluid interface [31]. The ratio between the reaction or incorporation flux and the mass transport flux can be represented by the Damköhler number (Da), which correlates transport phenomena, e.g. based on solutes, with time-dependent reaction rates [32] and facilitates the assessment of spatiotemporal distribution of the reaction rate under local non-equilibrium conditions as observed in precipitation/dissolution processes [33]. For NaCl precipitation from an aqueous solution during water evaporation, the Damköhler number was, for instance, shown to drastically change when the growth of sodium chloride crystals was observed [34]. Desarnaud et al. recently highlighted that in nature crystals are rarely found only in their equilibrium state [35]. Rather, many minerals and salts, like notably calcite and halite, form hopper crystals composed of clusters of interconnected crystalline regions. These authors point out that little fundamental work had been done on understanding hopper crystal shapes resulting from an anisotropic growth that is due to the edges of a crystal growing faster than the centers of its faces. With kinetics of growth being an important factor in determining the overall shape of a solid, a transition between cubic and hopper growth at a high supersaturation of around 1.5 was reported when the growth rate of the cubic crystal reached a maximum of around 7 µm/s [35]. Goto et al. presented dendritic morphologies of sodium chloride when grown in a thin gelatin gel matrix in which the mass transport is governed by diffusion. Furthermore, the authors demonstrated a significant influence of the [salt]/[gelatin] concentration ratio and the humidity around a drying aqueous solution on the halite crystal shape and growth velocity ranging from approximately 1 to 20 μm/s [11]. Choudhury et al. used a simple simulation algorithm incorporating aggregation and evaporation to reproduce the observed well-defined cross-over from a compact NaCl crystal morphology in aqueous gelatin to a dendritic pattern as water evaporation proceeded [10]. Lately, Yang et al. [36] reviewed self-assembly in hopper-shaped crystals and described the change of crystal morphology according to a change of the interfacial instability. They established the role of capping agents that may inhibit the adsorption of a newly added building block on the surface of a growing solid. With respect to crystalline evaporative deposition, McBride et al. [37] showed that the energetics between all the three phases involved in the crystallization of solutes, namely substrate, crystal, and liquid, contribute to the nucleation barrier, and the authors elucidated that salt solutions with lower nucleation barriers will pin-drop substrate/drop contact lines to form rings.

Based on these studies and recent reports [38], it becomes clear that exploiting the mutual influences between proteins and salts for material design is an approach that is gaining increasing technological relevance. Thus, it is important to gain a fundamental understanding of the protein/salt or peptide/salt interactions when solid materials are formed from an aqueous phase. Such knowledge on liquid-solid transitions in the presence of different salts will be of great relevance for controlling the formation and multifunctional properties of new protein-based composite materials. In natural and biotechnical processes, the respective solid formation often starts from a liquid formulation comprising a solvent, an inorganic solute, and an organic co-solute and proceeds via the formation of growth units [11] as relevant entities, their assembly into nuclei, and the growth of metastable or stable solid phases [17]. Alternatively, processes allocating

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inorganic and organic moieties from different formulations may be engineered, e.g. using layer-by-layer [39] or sol–gel processes [40]. Recently, within the scope of tailoring biomimetic hybrid collagen/chitosan hydrogels with a dual network structure [38], an immersion in a highly concentrated 5 wt% aqueous sodium sulfate solution was reported to result in an opaque gel during a universal soaking step following a UV-induced crosslinking step.

Based on knowledge from experimental or simulation approaches, target-oriented strategies can be framed not only for the synthesis of bio-inspired materials, e.g. nacrelike composites [41], or surfactants for interface engineering [42, 43], but also for assessing environmental, agricultural, alimentary, medical or pharmaceutical challenges [17, 19, 38, 42, 44–46], e.g. avoiding the formation of renal stones [44]. Systematic material development is significantly promoted by applying scale-comprehensive simulation, e.g. of growth processes [30, 47], and access to databases facilitated by data and metadata interoperability [48].

Lately, our group demonstrated how salt-driven self-assembly can be applied for the physiological preparation of nanofibrous fibrinogen scaffolds to support cell adhesion [49–51] and compared relevant approaches for fibringen fiber assembly in vitro [22]. By precipitating a water-soluble mixture of salts in the presence of fibrinogen, the salts enabled the formation of dense nanofiber networks upon drying. On the other hand, chloride salts of calcium, magnesium, copper and zinc hamper fibrinogen fiber formation [52]. Moreover, we routinely used highly concentrated salt solutions to assemble the extracellular matrix protein collagen into nanofibers to control the growth of different cell types [53, 54]. In view of the hierarchy for hybrid materials presented by Saveleva et al. [55], such self-assembly processes start from an organic molecule-modified inorganic material (organics-in-inorganics) and end with a potentially inorganic-modified organic material (inorganics-in-organics). In the present contribution, we therefore focus on revealing the interface-active action of the proteins BSA, fibrinogen and collagen type I in the organics-in-inorganics approach for directing the crystallization of the alkali metal salt sodium chloride upon drying protein-containing saline water. While BSA and fibringen are both blood plasma proteins, collagen contributes to the assembly of an extracellular matrix during the early stages of wound healing [56]. In detail, we present microscopic images for evaluating the crystal size distribution and energy dispersive x-ray analysis (EDX) studies for disclosing the presence of an organic surface film.

Materials and experimental procedure

Sample preparation

All solutions were prepared using ultrapure water from a TKA water purification system (Thermo Fisher Scientific, Schwerte, Germany). A sufficient amount of tris(hydroxymethyl)aminomethane (Tris, $\rm C_4H_{11}NO_3$, Carl Roth GmbH, Karlsruhe, Germany) to obtain a 10 mM concentration was dissolved in ultrapure water and the pH of the solution was adjusted to 7.4 with hydrochloric acid (aqueous HCl, VWR, Darmstadt, Germany). Likewise, sodium chloride (NaCl, VWR) was dissolved in the Tris–HCl buffer to obtain a stock solution concentration of 1.5 M. Fibrinogen stock solutions were prepared by dissolving 10 mg/mL fibrinogen (100% clottable, Merck, Darmstadt,

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Germany) in a 10 mM Tris-HCl solution. The fibrinogen stock solution was dialyzed against a 10 mM Tris-HCl solution overnight using cellulose membrane dialysis tubing with 14 kDa cut-off (Sigma, Steinheim, Germany) to remove low molecular weight compounds. The BSA stock solution was prepared by dissolving 10 mg/mL BSA (Sigma Aldrich Munich, Germany) in 10 mM Tris-HCl buffer. Collagen stock solutions were prepared by dissolving 10 mg/mL collagen type I from calf skin (Sigma Aldrich, Munich, Germany) in 5% aqueous acetic acid (Carl Roth GmbH, Karlsruhe, Germany). Round glass coverslips with a diameter of 12 mm (VWR, Darmstadt, Germany) were sputtercoated with a 5 nm thin adhesion layer of chrome, followed by 25 nm of gold, using an EM ACE600 high vacuum sputter coater (Leica Microsystems, Wetzlar, Germany). A volume of 70 µL of the respective protein solution was pipetted onto the gold-coated glass substrates, followed by the addition of 70 µL salt solution to the protein drop. For the preparation of samples without protein, 70 µL of the salt solution was pipetted onto the gold-coated glass followed by 70 µL of ultra-pure water to adjust the salt and Tris-HCl concentrations. The samples were placed in a container purged with dry N₂ gas, and after the droplet drying process the samples were transferred to a home-built humidity chamber to be incubated at 24 °C and 30% relative humidity overnight.

Microscopic surface analysis

All optical microscopy images were obtained with a Keyence VHX-7000 digital microscope (Keyence, Neu-Isenburg, Germany). The samples were imaged in overall survey applying stitching and using magnification from $20\times$ for single partial images. Details of the precipitated salt or the protein deposits were imaged with magnifications varying from $80\times$ to $500\times$. Moreover, after depositing a thin electrically conductive carbon layer more detailed inspection was performed with Scanning Electron Microscopy (SEM) using a field emission device (FESEM), type FEI Helios 600 (Dual Beam, FEI, Eindhoven, Netherlands). The resolution was 0.9 nm at 15 kV at optimal working distance and 1 nm at 15 kV at the coincidence point. The images of the sample surface were obtained at acceleration voltages between 0.35 and 30 kV and at working distances between 1 and

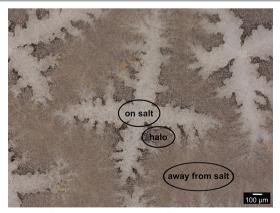


Fig. 1 Light microscopy image of deposits containing NaCl, fibrinogen and Tris moieties. Introduction of the terminology applied for surface areas imaged with higher magnification and SEM analysis: the surface of apparently crystalline salt deposits (labelled "on salt"), the halo close to the salt deposits (labelled "halo"), and regions beyond the halo (labelled "away from salt")

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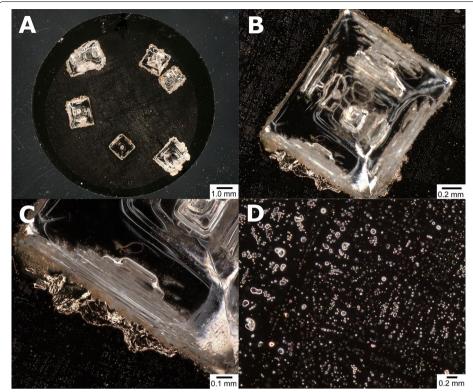


Fig. 2 Light microscopy images of NaCl crystals formed after drying a droplet of a solution containing 375 mM NaCl and 5 mM Tris-HCl buffer. (**A**) Overview image of the sample (×20 magnification), (**B**) selected NaCl crystal (×80 magnification), (**C**) detail of a top-sided edge of the selected crystal (×200 magnification) and (**D**) detail of deposits found away from the tabular crystals on the substrate surface (×500 magnification)

10 mm. For the detection of secondary or backscattered primary electrons an Everhart—Thornley or an in-lens detector was used.

EDX investigations

EDX measurements were performed with the same FESEM of type FEI Helios 600 (DualBeam) equipped with an Oxford X-Max80 silicon drift detector (SDD) for x-rays, an ATW2-window and an energy resolution down to 129 eV. The detection angle of the detector was 52°.

Results and discussion

The following discusses, the structure and arrangement of the NaCl crystals formed upon droplet drying as well as the composition of the deposits revealed in their surroundings. Hereby, the results of four scenarios are considered; namely the deposits resulting from water evaporation from Tris–HCl buffered aqueous formulations free of biopolymers in comparison with those obtained in the presence of BSA, fibrinogen or collagen, respectively. The Tris–HCl buffer was used to maintain the same physiological pH (7.4) for all four starting formulations involved in this study. In detail, three characteristic regional sections within the laterally inhomogeneous drying patterns were considered, as displayed in Fig. 1; these comprise the surface of apparently crystalline salt

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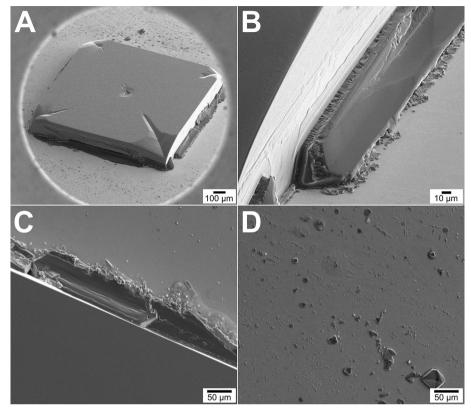


Fig. 3 SEM images of NaCl crystals formed after drying a droplet of a solution containing 375 mM NaCl and 5 mM Tris–HCl buffer. (**A**), (**B**) Show the SE-SEM images of an NaCl crystal and its halo region, respectively. (**C**) Displays the surface (bottom left) and halo regions (top right) of a salt crystal while (**D**) depicts the substrate area away from any NaCl crystals

deposits (called "on salt"), the halo close to the salt deposits (called "halo"), and regions beyond the halo (called "away from salt").

Macrostructure variation of NaCl crystals precipitated in the absence of proteins

After sodium chloride was dried in the absence of proteins the precipitated NaCl crystals were imaged with light microscopy and SEM. Figure 2 depicts the top view light microscopy images obtained after drying a droplet that was initially about 2 mm narrower than the substrate slide, as can be inferred from the approx. 1 mm wide dark rim shown in the light microscopic image in Fig. 2A. Water evaporation occurring over the course of two hours gave rise to the formation of six NaCl crystals with a rectangular, almost square base. While the width of these crystals varied between 1.3 and 2.5 mm, the heights of up to 0.2 mm were approximately one order of magnitude smaller. The tabular shape of the NaCl crystals was confirmed by the SEM in Fig. 3A, which illustrates that the crystal plateau height does not exceed the droplet height. Most of these crystals exhibited one edge that was less than 1 mm away from the initial contact line and the central substrate region was free from such distinct salt crystals. This finding can be explained by the observation that the decline of the liquid volume primarily resulted in a reduction of the droplet height rather than the droplet width. Details of the surface and the

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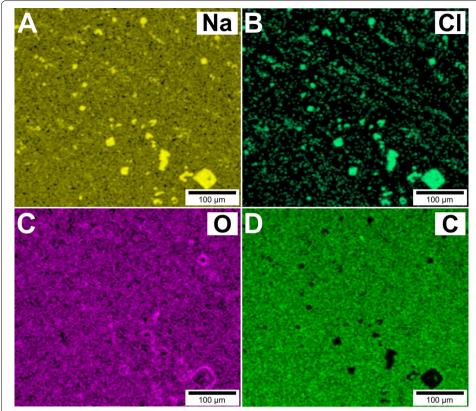


Fig. 4 EDX map false color representations of the substrate surface "away from salt", as shown in Fig. 3D obtained after drying a droplet of a solution containing 375 mM of NaCl and 5 mM Tris–HCl buffer: Na map (**A**), Cl map (**B**), O map (**C**), and C map (**D**)

shape of a 3D crystal can be seen in Fig. 2B, C. The crystal growth seemed to be faster at the crystal edges than at the crystal center or corners since steps and depressions were observed in the central region and at the corners of the crystals. The basal corners of the crystals show 90° angles, as expected for the growth morphology of halite, yet the topmost corners were not complete and exposed unfilled steps. In regions between the bigger crystals, and notably around the sample center, deposits of tiny NaCl crystals up to 30 µm in width can be observed as suggested by the light microscopy (see Fig. 2D) and SEM analyses (see Fig. 3D) and substantiated by the element contrast revealed in the EDX maps highlighting locally elevated Na and Cl concentrations (Fig. 4A, B). The bases of the halite crystals are bordered by amorphous deposits that are comparatively poor in Na or Cl species but rich in O and C containing moieties based on the contrast in the EDX maps shown in Figs. 4 and 5. Due to this elemental composition, these deposits can be attributed to solid Tris moieties.

As Tris-based deposits were observed and as they revealed a laterally inhomogeneous distribution we infer that Tris constitutes not only a buffer but also a co-solute of sodium chloride. Moreover, Taha et al. reported interactions between Na⁺ and Cl⁻ ions and Tris

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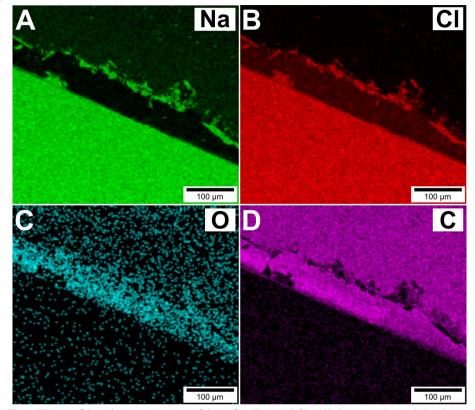


Fig. 5 EDX map false color representations of the surface (bottom left) and halo regions (top right) in the same region as shown in Fig. 3C for a NaCl crystal formed after drying a droplet of a solution containing 375 mM of NaCl and 5 mM tris–HCl buffer: Na map (**A**), Cl map (**B**), O map (**C**), and C map (**D**)

molecules [57]. The crystallization-related effects of the interactions between Tris, salt and water at interfaces are subject to ongoing studies.

Macrostructure variation of NaCl crystals precipitated in the presence of proteins

The following sections highlight the effects resulting from the presence of the proteins BSA, fibrinogen or collagen on the structure of sodium chloride precipitates as well as the arrangement and composition of organic deposits.

Effect of BSA

When 5 mg/mL BSA as a starting concentration was adjusted in the solution left for drying, seven bigger NaCl crystals with widths varying from 1 to 1.8 mm and a rectangular base were observed (Figs. 6A and 7A), similar to the salt precipitates obtained after drying the droplet without any protein. Similarly to Figs. 2B, C and 6B, C show more surface topography details of the central NaCl crystal. Figure 6D, in particular, shows the formation of more tiny crystals and dendrite growth from the edges towards the center of the substrate. However, the crystal shape formed in the presence of BSA was significantly different from NaCl deposits formed in the absence of protein since the bigger halite crystals were hoppered. In contrast, numerous arrangements of dendritic NaCl crystallites were found in regions away

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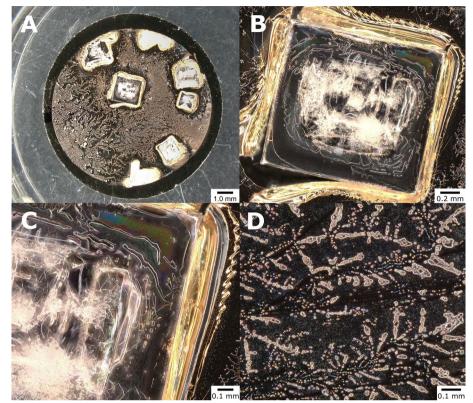


Fig. 6 Light microscopy of NaCl crystals formed after drying a droplet of a solution containing 375 mM of NaCl, 5 mM Tris–HCl buffer and 5 mg/mL BSA. (**A**) Overall image of the sample (\times 20 magnification), (**B**) close-up of the selected central NaCl crystal (\times 100 magnification), (**C**) detail of the top-edge of the selected crystal (\times 200 magnification), and (**D**) detail of deposits found away from the hoppered crystals on the substrate surface (\times 500 magnification)

from salt (see Figs. 6D and 7B-D), with their sizes varying between approximately 1 and 100 μ m. This finding indicates that the nucleation of salt crystals is not a limiting step within a nucleation and growth scenario. Rather, the observed multiplicity of small crystallites is attributed to a hindered transport of NaCl growth units in the BSA-containing fluid and, therefore, the dendrite formation may be inferred to occur in a later stage of the droplet drying than the growth of huge hoppered halite crystals. As such prevalence of salt dendrites was not observed upon the drying of protein-free droplets, we suggest that in this ultimate drying stage the viscosity of the remaining liquid was higher due to the presence of BSA and its self-assembly by the increase in salt concentration. Previously, similar aspects were addressed with respect to drying aqueous gelatin formulations resulting in a viscous gel film upon water evaporation [10, 58]. Such a protein-related effect may be in addition to the effect experienced by the salt growth units through the successively increasing salt concentration upon water evaporation [59].

As revealed by light microscopy, SEM analysis and the EDX maps obtained from the surface of halite hoppers (cf. Figs. 6B, C and 8), the central hopper region acted

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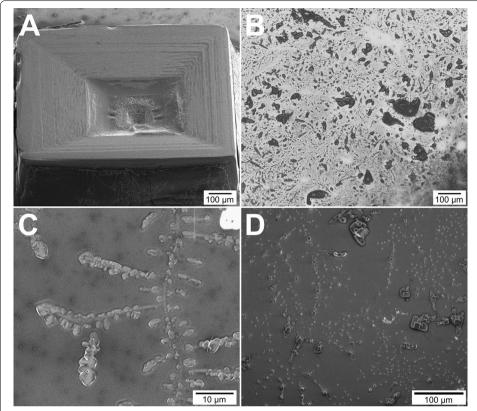


Fig. 7 SEM images obtained after drying a droplet of a solution containing 375 mM NaCl, 5 mM Tris–HCl buffer and 5 mg/mL BSA. (**A**) SE-SEM image of a hoppered NaCl crystal, (**B**) BSE-SEM image of a detail close to the halo of a NaCl crystal, (**C**) SE-SEM image obtained in the halo, and (**D**) overview image away from salt

as a pool for the remaining liquid during droplet drying. The thus provided microenvironment featured deposits that were dominated by Tris and its hydrochloride (Tris–HCl), as can be concluded from the EDX findings listed in Table 1. In detail, the corresponding EDX maps not only showed a significant atomic concentration of chloride without revealing any metal cations but also revealed an [N]/[O] concentration ratio around 0.45, which is similar to the stoichiometrically expected ratio for Tris, i.e. [N]/[O] = 0.33, or possibly mixtures resulting from interactions of Tris with BSA [60], which are expected to be closer to 1 in view of the predominant ratio of [N]/[O] = 1 of the peptide groups in proteins. Moreover, the organic deposits visible via light microscopy and SEM seem to decorate and wrap the salt hoppers as well as the smaller crystallites around their borders (see Figs. 6B, 7B and 9C).

Effect of fibrinogen

When 5 mg/mL fibrinogen as a starting concentration was added to the NaCl solution and left to drying, no apparently single crystalline halite deposits more than 1 mm wide were observed. Rather, the original contact line was decorated by a rim of NaCl deposits less than 0.5 mm wide. Around the initial droplet center, a dozen crosswise salt dendrites with maximum lengths between 1 and 2 mm and maximum

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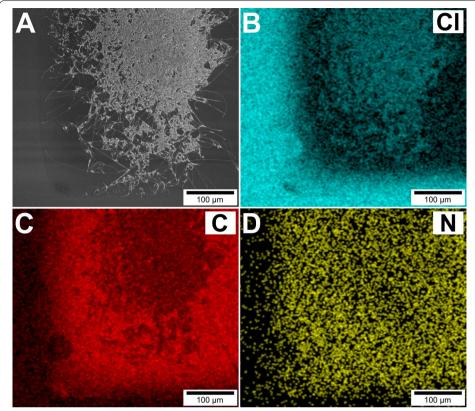


Fig. 8 SEM image and EDX map false color representations in the same region of the salt surface obtained after drying a droplet of a solution containing 375 mM NaCl, 5 mM Tris–HCl buffer and 5 mg/mL BSA: (**A**) SEM image, (**B**) Cl map, (**C**) C map, and (**D**) N map

Table 1 Evaluation of the EDX investigations for distinct surface regions of deposits from NaCl with and without different proteins obtained from Tris-buffered droplets

Sample	[C] (at%)	[N] (at%)	[O] (at%)	[Na] (at%)	[CI] (at%)	[S] (at%)	[N]/[O]
Surface of tabular NaCl crystal, without protein (10 kV)	21.1	_	0.3	40.2	38.4	-	=
Surface of hoppered NaCl crystal, with BSA (10 kV) like in Fig. 8	56.2	12.2	27.0	=	4.6	=	0.45
Beading in halo of hoppered NaCl crystal, with BSA (10 kV)	57.7	18.5	23.3	0.3	0.9	0.3	0.79
Overall dendritic NaCl crystal and halo, with fibrinogen (10 kV)	61.6	6.5	7.1	12.9	11.7	0.1	0.92
Around center of dendritic NaCl crystal, with fibrinogen (10 kV)	53.7	3.6	2.9	20.2	19.4	0.1	1.2
Away from dendritic NaCl crystal, with fibrinogen (10 kV)	69.4	8.4	8.8	6.9	6.0	0.2	0.95
Overall dendritic NaCl crystal and surrounding, with collagen (10 kV) like in Fig. 14B	59.8	12.1	22.4	2.5	3.2	0.05	0.54
Tris, according to stoichiometric composition	50	12.5	37.5	_	_		0.33

The atomic concentrations (at%) and the [N]/[O] concentration ratio are given as obtained with an electron acceleration voltage of 10 kV

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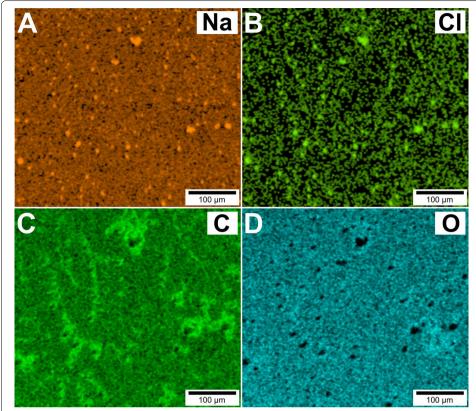


Fig. 9 EDX map false color representations in the region "away from salt" as displayed in Fig. 7D and obtained after drying a droplet of a solution containing 375 mM NaCl, 5 mM Tris–HCl buffer and 5 mg/mL BSA: Na map (**A**), Cl map (**B**), C map (**C**), and O map (**D**)

widths between 0.2 and 0.3 mm were observed (see Fig. 10A). The compact packing within the manifestly polycrystalline dendritic NaCl deposits (see Fig. 11C) indicates that the heterogeneous nucleation of the up to several micrometer wide cubic grains on the surface took place, while the lateral growth stopped at the grain boundaries. The near-surface material contrast achieved by the detection of back-scattered electrons (BSE) as shown in Fig. 11B, C and the elemental distribution revealed by the EDX maps (see Fig. 12) recorded on top and around the intersection area of the two orthogonal branches constituting the crosswise NaCl dendrite revealed three distinct characteristics. First, the elevated regions of the salt deposits are covered by patches of an organic substance composed of elements with a lower atomic number than Na or Cl. The findings from the EDX analysis listed in Table 1 reveal a significant attenuation of Na and Cl signals from the underlying sodium chloride and an [N]/[O] atomic concentration ratio around 1. Correspondingly, the organic deposits are interpreted to be predominantly composed of fibringen multilayers. Second, an approximately 100 µm wide halo that is depleted in NaCl is visible close to the central intersection area of the crosswise dendritic crystals. Third, in comparison to the central dendrite regions, the lower extensions adjacent to the principal branch revealed a lower coverage by such organic moieties.

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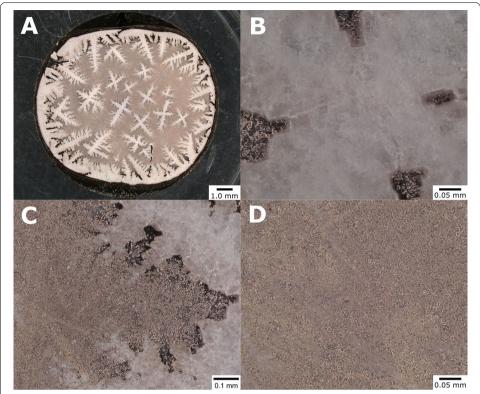


Fig. 10 Light microscopy of NaCl crystals formed after drying a droplet of a solution containing 375 mM NaCl, 5 mM Tris–HCl buffer and 5 mg/mL fibrinogen. (**A**) Overview image of the sample (×20 magnification), (**B**) detail on salt (×100 magnification), (**C**) detail on the halo region (×200 magnification) and (**D**) detail of deposits found away from the bigger crystals on the substrate surface (×500 magnification)

Since we did not detect any noticeable X-ray emission from the underlying gold substrate we conclude that the respective NaCl moieties were deposited on top of an organic film. This film is thicker than that on top of the NaCl dendrites. Similar findings were obtained in regions away from the salt dendrites, as shown in Fig. 11D and in Table 1, which once again revealed an [N]/[O] atomic concentration ratio of around 1. In these regions, NaCl crystallites a few micrometer wide and with a cubic or cubo-octahedral shape due to exposed {100} or partially {111} faces were observed (Fig. 11D). Therefore, we suggest that the halite formation contributing to the aforementioned extensions and the deposits in the last-mentioned regions proceeded after the precipitation of the principal NaCl branch in the ultimate phase of the droplet drying process.

Effect of collagen

The final investigation focused on the precipitates formed when drying droplets containing 5 mg/mL collagen as a starting concentration. The findings obtained by light microscopy (see Fig. 13), SEM (see Fig. 14), and EDX mapping (see Fig. 15) revealed the formation of polycrystalline crosswise dendrites extending from the initial contact line towards the sample center. Similarly, orthogonal dendrites with straight branches of sodium chloride were observed when grown in a thin gel matrix made of gelatin

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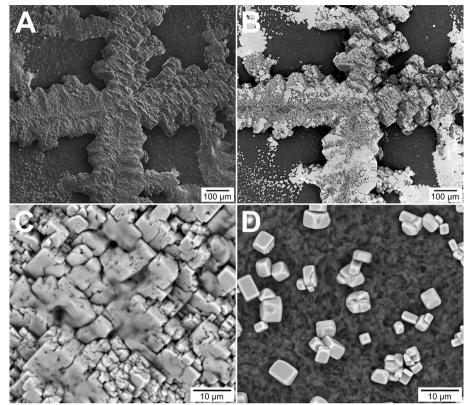


Fig. 11 SEM images obtained after drying a droplet of a solution containing 375 mM NaCl, 5 mM Tris–HCl buffer and 5 mg/mL fibrinogen: details on top and around NaCl dendrites, as contrasted detecting (**A**) secondary (SE) or (**B**) back-scattered electrons (BSE). (**C**) BSE-SEM views of the dendrite surface, (**D**) BSE-SEM images of the region away from the dendrites

[11], a collagen hydrolysate, at mass concentration ratios [gelatin]/[NaCl] of around 0.5; meanwhile, we used a mass concentration ratio of 0.23 in the Tris buffer. In the dendrites, the sizes of the elevated halite grains on top of the approximately 0.2 mm high central NaCl crystallite ranged from 30 to 50 μ m (see Fig. 14A–C), meaning they were significantly bigger than the ones obtained in the presence of fibrinogen.

Effects of distinct proteins in direct comparison

By comparing the microscopic findings for the structure and arrangement of halite crystallites formed during the drying of droplets containing aqueous formulations buffered with 5 mM Tris and a given NaCl concentration of 22 mg/mL we found that adding an initial 5 mg/mL concentration of a protein chosen among BSA, fibrinogen or collagen significantly and specifically changed the obtained drying pattern on gold-coated glass slides. On the one hand, the addition of the globular protein BSA did not strongly affect the overall drying pattern and—as also observed in the absence of proteins—still gave rise to the formation of approximately 1 mm wide halite crystals with a clear and transparent optical appearance. However, in the presence of BSA, the crystal shape was not tabular as in the absence of any protein, but hoppered. On the

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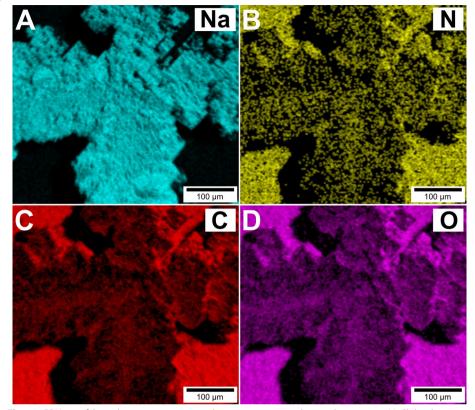


Fig. 12 EDX map false color representations in the region on top and around a crosswise NaCl dendrite as obtained after drying a droplet of a solution containing 375 mM NaCl, 5 mM Tris–HCl buffer and 5 mg/mL BSA: **(A)** Na map, **(B)** N map, **(C)** C map, and **(D)** O map

other hand, adding the fibrillar proteins fibringen or collagen resulted in the formation of polycrystalline dendritic NaCl deposits with halite grain sizes clearly below 0.1 mm and an opaque optical appearance. Hence, the very nature of a specific protein and its complex interactions with the co-solutes NaCl and Tris significantly govern the formation of halite grain boundaries. We suppose that in the case of fibrinogen and collagen, the resulting salt crystals are shaped by persisting interphases between the salt and the circumjacent aqueous formulation, impedinge an Ostwald ripening of neighboring halite crystallite grains over distances exceeding 0.1 mm. Facilitating such longer-range transport in the liquid phase or even water monolayers [61] would be expected to result in a reduction of the number of smaller sodium chloride deposits for the benefit of bigger crystals with an energetically more favorable faceting of their habits. We thus rather infer that the rate of solute transport may be affected by the viscosity of the concentrated mobile phase, while the restructuring of the halite grain surfaces by the loss and gain of growth units may be influenced by protein-containing films and adsorbates. In this way, both the transport and the surface-related reaction term contributing to the Damköhler number characterizing the dynamics between the stationary and mobile phases will depend on the presence of a specific protein.

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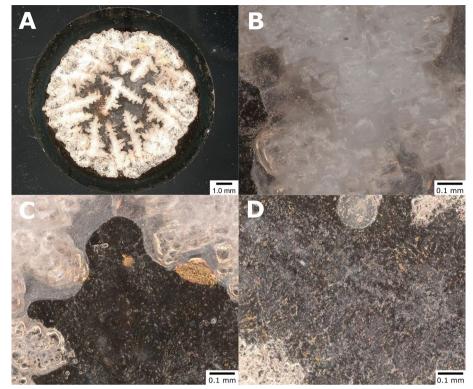


Fig. 13 Light microscopy of NaCl crystals formed after drying a droplet of a solution containing 375 mM NaCl, 5 mM Tris–HCl buffer, 375 mM acetic acid and 5 mg/mL collagen. (**A**) Overview image of the sample (×20 magnification), (**B**) detail of an "on salt" region (×300 magnification), (**C**) detail of the halo region (×200 magnification) and (**D**) detail of the region away from the salt (×500 magnification)

Based on our findings, we suggest that the specific protein-NaCl interactions are guided by the behaviour of the respective protein at the salt/liquid interphase, with the effect of collagen being rather more similar to that of fibrinogen than to that of BSA. Globular proteins on the one hand, and fibrous proteins, on the other hand, not only differ in the principles of their topology but also in the relative abundance of apolar and polar side chains in the amino acids constituting these proteins and the molecular flexibility [62]. For example, Cacace et al. highlighted that at pH 7 in the hydrated state the surface tension of human serum albumin (HSA) features a lower contribution from apolar Lifschitz-van der Waals interactions and a higher share of polar electron acceptor and electron donor potentials than the surface tension of fibrinogen. Therefore, we hypothesize that the specific interactions between a protein and the halite surface significantly influence the observed drying pattern. In detail, the reaction-related contributions to adhesion development within the frame of the macrokinetic approach based on a characteristic Damköhler number may be manifold. The process steps governing the formation of the interphase between the growing solid phase, e.g. salt crystals or solidified protein, and the highly viscous saline aqueous jelly phase containing protein may be related to two principal reaction-type binding events [63]: firstly, the often exothermic binding between cations or anions and the solid phase and, secondly, the entropy-driven and often endothermic water

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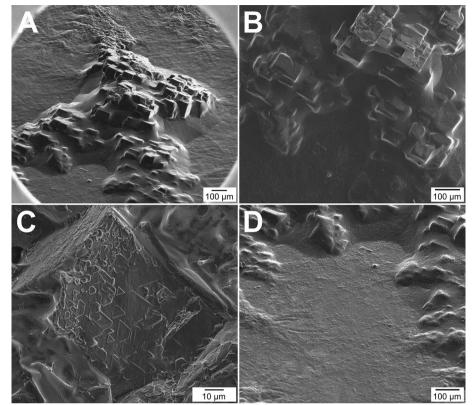


Fig. 14 SEM images of NaCl dendrites formed after drying a droplet of a solution containing 375 mM NaCl, 5 mM Tris–HCl buffer, 375 mM acetic acid and 5 mg/mL collagen. (**A**) Shows a tilted (52°) view of NaCl dendrites ("on salt" region), (**B**), (**C**): top-view details of different crystallites, and (**D**) region between dendritic NaCl crystals

replacement—e.g. upon dehydration of inorganic or organic moieties—in interphases, such as at highly dynamic salt crystal or protein interfaces in contact with the fluid environment.

In this way, acquiring knowledge on the formation of hybrid or composite materials in droplets of saturated aqueous solutions composed of several co-solutes in environments involving salts and proteins may be inspired by nature and promote medical, pharmaceutical or technological innovation. As highlighted by Lohse and Zhang [64], the spatiotemporal development of multicomponent fluid dynamical systems out of equilibrium is governed by concentration gradients inducing a transport concurring with phase transitions. These complex and interwoven phenomena and the challenges and opportunities related to them may be dealt with by the experimental, numerical and methodical tools and procedures provided by implementing the digital transformation not only in industry but also in research and development. Hence, exploring and designing the spatiotemporal evolution of adhesion is a topic that shows great promise in profiting from advanced materials modelling.

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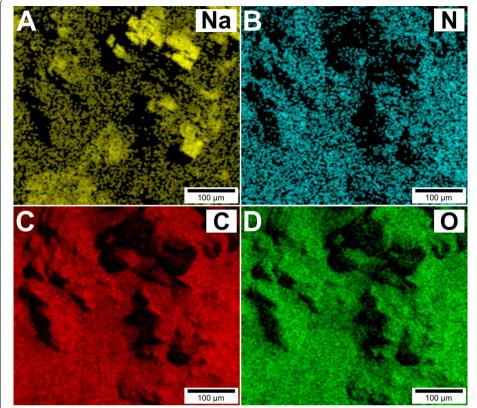


Fig. 15 EDX map false color representations in the region 375 mM NaCl, 5 mM Tris–HCl buffer, 375 mM acetic acid and 5 mg/mL collagen: (**A**) Na map, (**B**) N map, (**C**) C map, and (**D**) O map

Conclusions

In this study on the drying patterns formed from aqueous saline formulations, we found that the NaCl crystal arrangements and morphologies varied with the composition of the initial solution. Depending on the type of the protein chosen, among BSA, fibrinogen and collagen, the halite deposits predominantly exhibited cubic or polycrystalline dendritic structures. Based on these results, we hypothesize that the process steps in the formation of the interphase between the growing solid phase and the highly viscous saline aqueous jelly phase containing protein govern not only the material transport in the liquid but also the material exchange between solid and liquid phases. In the future, the synthesis of bio-inspired, protein-based composite materials will benefit immensely from understanding fundamental solid-liquid phase interactions to successively tailor these scale-comprehensive dynamic processes.

Abbreviations

BSA: Bovine serum albumin; Tris: Tris(hydroxymethyl)aminomethane; Da: Damköhler number; EDX: Energy dispersive X-ray analysis; SEM: Scanning electron microscopy.

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Authors' contributions

StS prepared the samples and set up the light microscopy experiments supported by KB and VCB, and contributed to the lay-out, drafting and formatting of the article; KB and VCB also contributed to the discussion of microscopic characterization; KT performed the SEM and EDX measurements and contributed to the discussion of microscopic characterization; WLC and MN took part in defining and setting up the experiments, discussing and merging the obtained data and in drafting the manuscript; IG, PS and SD took part in defining and setting up the experiments based on emerging material design challenges; DB contributed in planning the conceptual approach for joint research and in discussing results and drafting the manuscript. All authors read and approved the final manuscript.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author upon reasonable request.

Declarations

Competing interests

The authors declare that they have no competing interests.

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