

## EUGLENOIDS FROM THE EL FARAFRA OASIS (WESTERN DESERT, EGYPT)

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**Abstract.** The paper reports the first detailed study of the diversity of euglenoids recorded from the El Farafrā Oasis in the Western Desert of Egypt. Four of the 20 identified species are new records for the Egyptian algal flora: the colorless *Peranema inflexum* Skuja, and the three pigmented species *Euglena adhaerens* Matv., *Phacus crassus* Zakryś & M. Łukomska and *Ph. cristatus* Zakryś & M. Łukomska. A brief description and original documentation are given for each reported morphospecies.

**Key words:** Africa, biodiversity, El-Farafrā Oasis, Egypt, Euglenophyta, new records

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### INTRODUCTION

About a third of the world's land surface has arid or semi-arid climate. Africa is one of the continents with the most dry land. In the northern part of the continent is the largest desert in the world, the Sahara. Oases are a characteristic element of it. Oases are groundwater-sustained islands of verdant fertility in a barren and relatively isolated landscape (Powell *et al.* 2015; Powell & Fensham 2016).

On the basis of the *ca* 2100 records in Levants and Resburg's (2010) bibliography on the freshwater algae of Africa, which includes most of the available literature, we may conclude that the African freshwater algal flora is not sufficiently investigated, particularly in the northeastern region. Wołowski (2012) pointed out that most of the information about the biodiversity of African euglenoids relates mainly to East and Central Africa and narrow strips of Northern and Southern Africa. Recent literature offers a relatively good background on the

phycological structure of other algal and cyanobacterial groups in African countries (e.g., Touliabah *et al.* 2002; Shaaban *et al.* 2013, 2015; Amarouche-Yala *et al.* 2014; Bere & Mangadze 2014; Edger *et al.* 2015; Mansour *et al.* 2015; Cantonati *et al.* 2016; Saber & Cantonati 2016; Saber *et al.* 2016; Taylor *et al.* 2016; Janse van Vuuren & Taylor 2016). Previous studies of euglenoids reported a preponderance of common and cosmopolitan taxa that are considered good bioindicators for eutrophic and polluted inland waters, even those recorded from extreme, isolated or remote habitats. To complete our knowledge of the euglenoids of Africa, however, more in-depth studies are still needed, particularly in oases and other remote habitats. To date only a few papers have been devoted to the euglenoids of Africa (van Oye 1924, 1927; Conrad & Van Meel 1952; Gerrath & Denny 1979; Couté & Iltis 1981; Kadiri 1992, 2004; Kadiri & Opute 2000; Kemka *et al.* 2004; Zongo *et al.* 2006; Nguetsop *et al.* 2007; Da *et al.* 2009; Wołowski 2012). Compère's (1975) studies on Lake Chad and its

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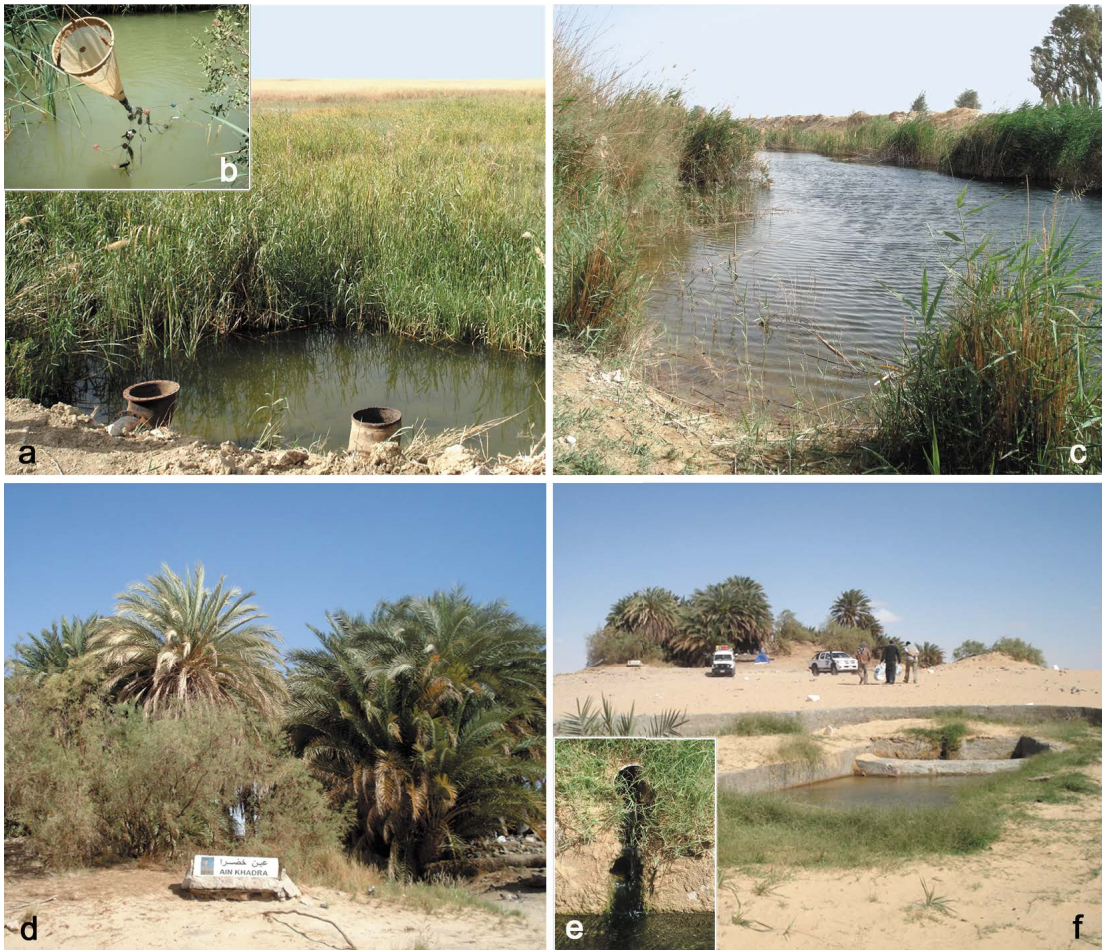
## MATERIALS AND METHODS

### STUDY SITE

El-Farafra Oasis (26°00'–27°30'N, 26°30'–29°00'E), located *ca* 650 km southwest of Cairo, is one of the smallest oases (*ca* 10,000 km<sup>2</sup>) on the limestone plateau occupying the central part of western Egypt (Fig. 1) (El Bastawesy & Ali 2013). This natural depression is in a hyper-arid region with a hot desert climate; it has 10 mm or less average annual precipitation and *ca* 22°C mean annual air temperature (MAAT) (Elsheikh 2015; Powell & Fensham 2016). Groundwater in the El-Farafra Oasis is drawn mainly from natural and artificial wells discharging from the world's largest

non-renewable groundwater resource, the Nubian Sandstone Aquifer (Voss & Soliman 2014). The specimens in this study were collected mainly from three different habitats in the oasis (Figs 1 & 2): (1) agricultural-water-fed Abu Nuss Lake (27°09'49.4"N, 27°55'46.2"E, 60 m a.s.l.) in El-Nahda village, (2) an agricultural drainage (27°03'30.5"N, 27°54'03.3"E, 70 m a.s.l.) in Lewa Soubah village, and (3) a slow-flowing rheocrenic ambient spring, Ain Khadra, also called Ain El-Wadi (27°22'15"N, 28°13'08.8"E, 31 m a.s.l.) in White Desert National Park (WDNP).

Abu Nuss Lake is one of the main attractions of the El-Farafra Oasis, and is home to abundant birdlife (Fig. 2a). Located in El-Nahda village, it has a total area



**Fig. 2.** Landscape views of the sampling sites: a – Abu Nuss Lake with massive growth of the common reed *Phragmites australis* (Cav.) Trin. ex Steud., b – plankton net hooked by its cone with a fishnet in Abu Nuss Lake, c – agricultural drainage at Lewa Soubah village, d – springhead of Ain Khadra (sustaining vegetated hillocks of palm trees), e – water flowing from Ain Khadra through an underground polypropylene tube, f – small concrete pool receiving water from Ain Khadra.

of *ca* 15 km<sup>2</sup> and ranges in depth between 2 and 3 m. It is *ca* 11 km north of Qasr El-Farafra, the capital and administrative center of El-Farafra Oasis. This lake might be more properly classified as a wetland, and is mainly fed by the water discharged from neighboring agricultural land. It is widely used for fish farming (Fig. 2b). The lake is densely covered with massive growth of the common reed *Phragmites australis* (Cav.) Trin ex Steud.

The second studied habitat is an agricultural drainage in the vicinity of Lewa Soubah village. The water of this shallow drainage (mean depth *ca* 1.5 m) is supplied by runoff from agricultural land (Fig. 2c).

The third investigated station, a rheocrenic ambient spring called Ain Khadra, is one of the three main springs that remained active in the WDNP. It is located in the northern extremity of El-Farafra Oasis, *ca* 35 km from Qasr El-Farafra. This spring still yields freshwater, with average discharge of 7 L min<sup>-1</sup> (Powell & Fensham 2016), and sustains vegetated hillocks rising several meters above the plain (Fig. 2d). The springhead is completely shaded and discharges its water through

a large polypropylene tube (Fig. 2e) in a small concrete pool used for recreation and for animals to drink in this isolated desert habitat (Fig. 2f).

HYDROCHEMICAL CHARACTERIZATION

Water was sampled using polyethylene bottles previously cleaned with ultrapure water and Suprapure nitric acid (1%). Water temperature (°C), pH, specific conductivity (μS cm<sup>-1</sup>) and total dissolved solids (mg l<sup>-1</sup>) were measured *in situ* using a HANNA HI 991301 calibrated portable Temp/pH/EC/Total Dissolved Solids (TDS) meter. Dissolved oxygen (DO) was determined with a Lutron® YK-22DO in-field calibrated dissolved oxygen meter. Detailed hydrochemical characteristics of the studied habitats, including major ions, nutrients, trace elements and metals were determined by standard procedures and methods (Clescerl *et al.* 2001). Metals were analyzed with a PerkinElmer Optima 5300 inductively coupled plasma optical emission spectrometer (ICP-OES). Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>2+</sup> and Mg<sup>2+</sup> were measured by ionic chromatography (ICS 1500, Dionex Corp.).

**Table 1.** Main hydrochemical characteristics of the studied habitats in El-Farafra Oasis (Western Desert, Egypt).

| Parameter                                           | Abu Nuss Lake<br>in El-Nahda village | Agricultural drainage<br>in Lewa Soubah village | Ain Khadra |
|-----------------------------------------------------|--------------------------------------|-------------------------------------------------|------------|
| Temp. (°C)                                          | 24.5                                 | 22.4                                            | 24.0       |
| pH                                                  | 7.3                                  | 6.6                                             | 7.25       |
| Conductivity (μS cm <sup>-1</sup> )                 | 4040                                 | 5930                                            | 330        |
| T.D.S. (mg l <sup>-1</sup> )                        | 2090                                 | 2990                                            | 170        |
| Dissolved oxygen (mg l <sup>-1</sup> )              | 6.2                                  | 4.4                                             | 1.5        |
| Na <sup>+</sup> (mg l <sup>-1</sup> )               | 502                                  | 820                                             | 14.5       |
| K <sup>+</sup> (mg l <sup>-1</sup> )                | 49.3                                 | 62.2                                            | 20.2       |
| Ca <sup>2+</sup> (mg l <sup>-1</sup> )              | 145                                  | 243                                             | 10.8       |
| Mg <sup>2+</sup> (mg l <sup>-1</sup> )              | 128                                  | 172                                             | 14.2       |
| Cl <sup>-</sup> (mg l <sup>-1</sup> )               | 944.5                                | 2119.32                                         | 102.03     |
| SO <sub>4</sub> <sup>2-</sup> (mg l <sup>-1</sup> ) | 532.7                                | 343.63                                          | 1.11       |
| HCO <sub>3</sub> <sup>-</sup> (mg l <sup>-1</sup> ) | 163.71                               | 100                                             | 24.37      |
| CO <sub>3</sub> <sup>2-</sup> (mg l <sup>-1</sup> ) | 0.0                                  | 0.0                                             | 0.0        |
| NO <sub>2</sub> <sup>-</sup> (μg l <sup>-1</sup> )  | 112                                  | 452                                             | 0.0        |
| NO <sub>3</sub> <sup>-</sup> (μg l <sup>-1</sup> )  | 3760                                 | 157000                                          | 400        |
| NH <sub>4</sub> <sup>+</sup> (μg l <sup>-1</sup> )  | 60                                   | 80                                              | 15         |
| TP (μg l <sup>-1</sup> )                            | 325                                  | 0.0                                             | 0.0        |
| SiO <sub>2</sub> (mg l <sup>-1</sup> )              | 6.8                                  | 9.2                                             | 5.5        |
| Sr (μg l <sup>-1</sup> )                            | 3497                                 | 5860                                            | 233        |
| B (μg l <sup>-1</sup> )                             | 68.6                                 | 64.6                                            | 137        |
| Li (μg l <sup>-1</sup> )                            | 49.1                                 | 79.6                                            | 4.0        |
| Fe (μg l <sup>-1</sup> )                            | 32.3                                 | 24.0                                            | 29.4       |
| Mn (μg l <sup>-1</sup> )                            | 5.1                                  | 2.9                                             | 4.6        |
| Zn (μg l <sup>-1</sup> )                            | 3.2                                  | 3.0                                             | 7.6        |
| Cu (μg l <sup>-1</sup> )                            | 0.5                                  | 0.84                                            | 0.12       |

HCO<sub>3</sub><sup>-</sup>, CO<sub>3</sub><sup>2-</sup>, Cl<sup>-</sup> and SO<sub>4</sub><sup>2-</sup> were determined according to Chapman and Pratt (1978). Nutrients (NO<sub>2</sub><sup>-</sup>, NO<sub>3</sub><sup>-</sup>, NH<sub>4</sub><sup>+</sup>, TP) were measured by molecular absorption spectrometry, and silicates as SiO<sub>2</sub> by the molybdsilicate method (Clescerl *et al.* 2001). The main hydrochemical characteristics of the studied habitats in El-Farafra Oasis are given in Table 1.

COLLECTED MATERIALS

Euglenoid specimens were sampled April 9–11 2015. The specimens were filtered using a plankton net (15 µm mesh) and transferred in 100 ml sterile clean polyethylene bottles following the method adopted by Bellinger and Sigeo (2010). Fresh specimens were transported chilled in a cooler to the laboratory. The materials were studied and identified using a BEL® Photonics biological light microscope, and later preserved with 4% (v/v) formaldehyde solution. All descriptions are based on morphotaxonomic observations of living specimens. Voucher specimens were deposited in Phycology Unit No. 341, Botany Department, Faculty of Science, Ain Shams University, Cairo, Egypt. Diagnostic morphometric characters were measured and photographed using Canon Powershot G12 digital camera; 15 measurements of each morphometric character were made. The specimens were identified using the relevant literature: Starmach (1983), Wołowski (1998), Wołowski and Hindák (2005) and Wołowski (2011).

RESULTS AND DISCUSSION

In the studied habitats of the El-Farafra Oasis we recorded 20 different species and intraspecific taxa of euglenoids belonging to 8 genera: *Peranema* Dujard. (1), *Euglena* Ehrenb. (4),

*Euglenaformis* M. S. Bennett & Triemer (1), *Euglenaria* Karnkowska-Ishikawa, E. W. Linton & Kwiat. (2), *Discoplastis* Triemer (1), *Lepocinclis* Perty (4), *Phacus* Dujard. (6) and *Trachelomonas* Ehrenb. (1) (Table 2). All these taxa have a cosmopolitan distribution and are known to thrive in moderately to highly polluted waters rich in organic matter (Wołowski 2011; Duangjan *et al.* 2014; Triemer & Zakryś 2015). Our results confirm that these euglenoids usually prefer, and proliferate in, warm and stagnant waterbodies (e.g., Wołowski 1998, Ciugulea & Triemer 2010; Wołowski *et al.* 2013; Shaaban *et al.* 2015; Varol & Şen 2016). These habitat conditions are observed mainly in two of the three studied habitats: Abu Nuss Lake and the agricultural drainage, which showed high concentrations of N and P compounds (see Table 1 for more details). Only *Trachelomonas volvocina* Ehrenb. var. *volvocina* was found in the rheocrenic ambient spring known as Ain Khadra. Shaaban and El Habibi (1978) documented similar findings in his study of some drilled wells in the El-Kharga Oasis, where only *T. hispida* (Perty) F. Stein and *T. volvocina* were recorded. Wołowski (1998, 2011) and Poniewozik (2009) reported that the genus *Trachelomonas* Ehrenb. mostly prefers colder and clean waters, although it can also be found in warmer and polluted waters.

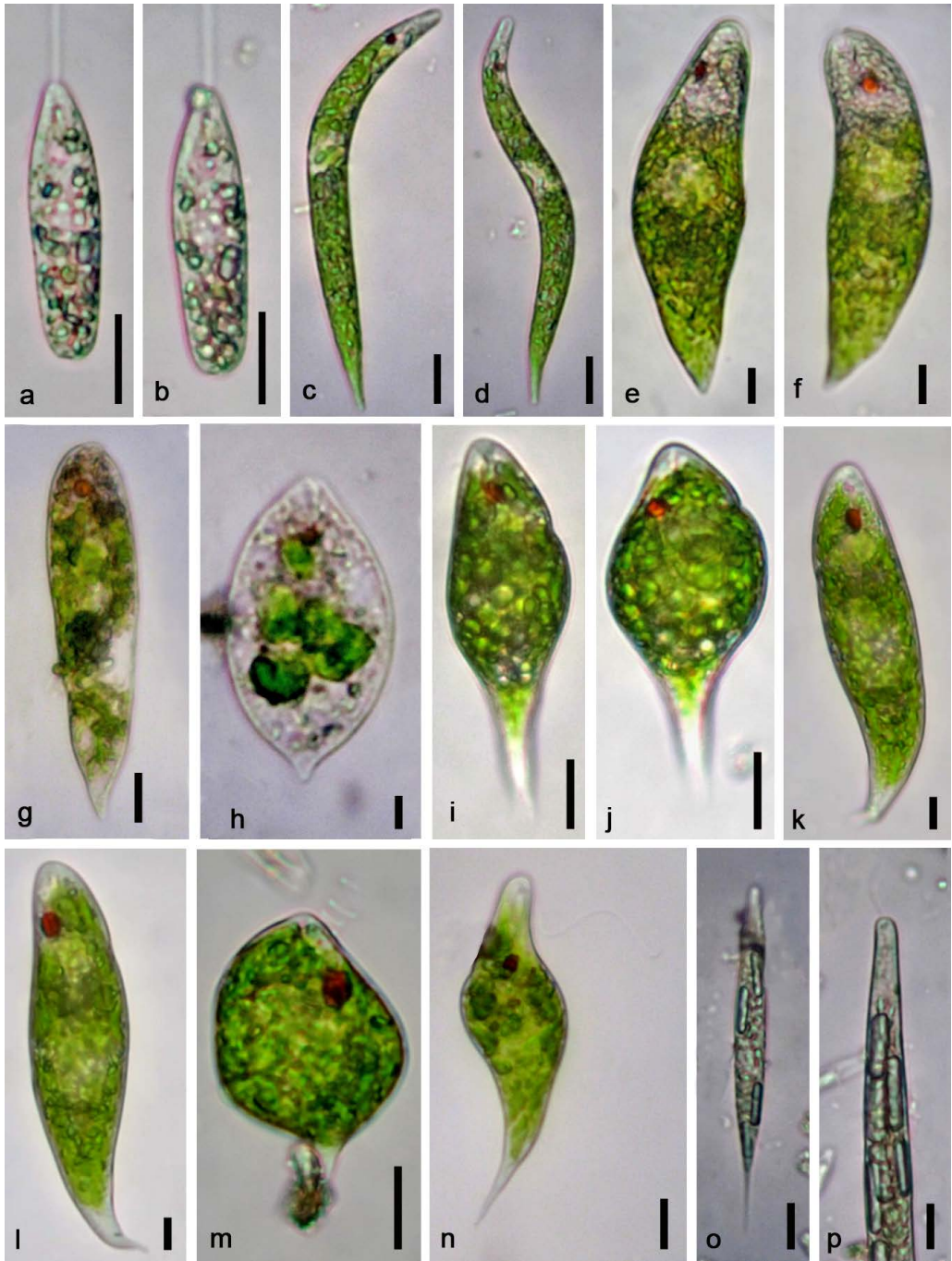
*Phacus crassus* Zakryś & M. Łukomska and *Ph. cristatus* Zakryś & M. Łukomska were previously included in the species complex *Ph. longicauda* in assessments of Egyptian euglenoids

**Table 2.** Euglenoid taxa reported from various sites of El-Farafra Oasis (I – Abu Nuss Lake, II – agricultural drainage, III – Ain Khadra ambient spring).

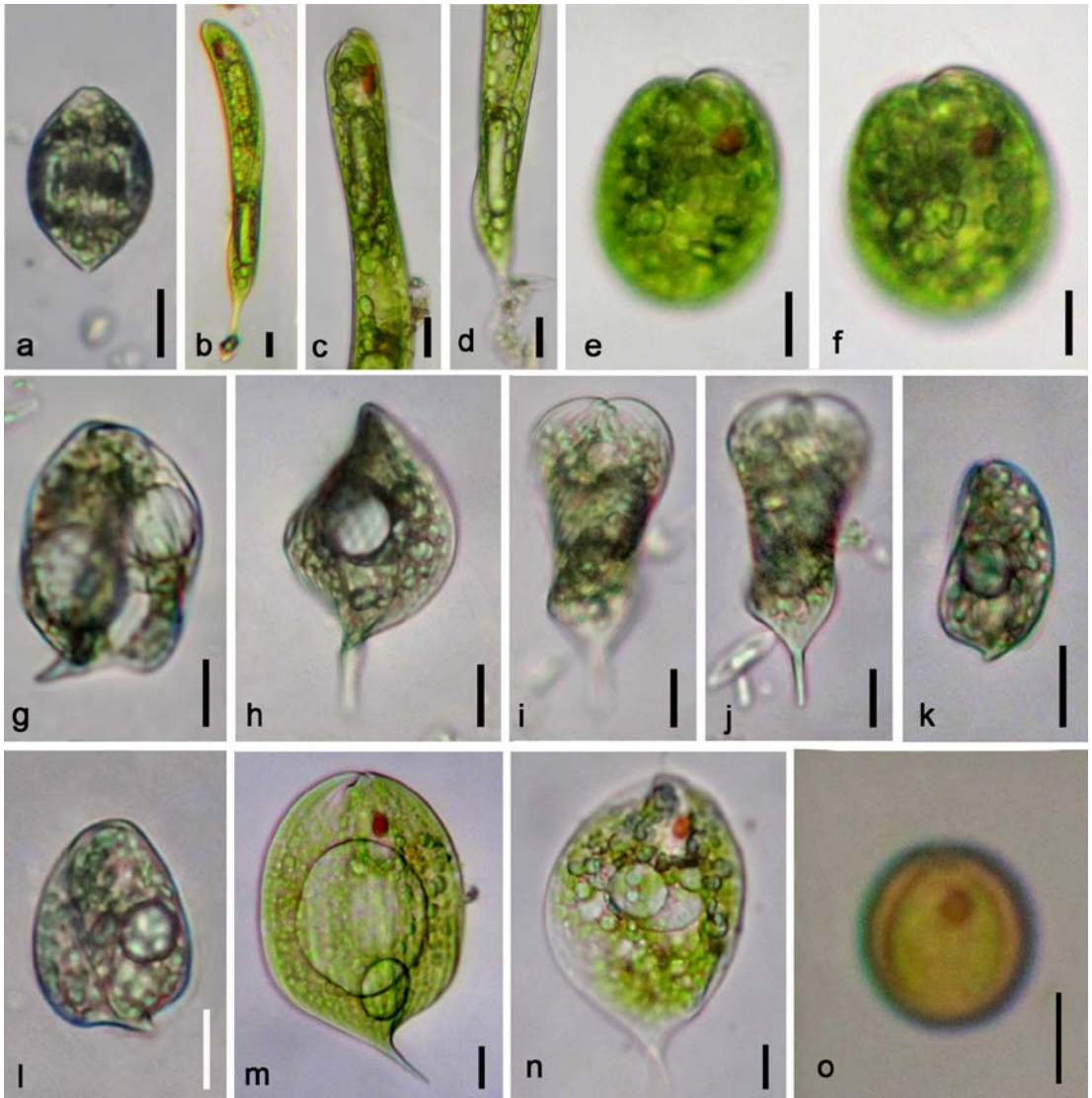
| Figs  | Taxon                                        | Taxonomical descriptions                                                                                             | Sites and occurrence |    |     |
|-------|----------------------------------------------|----------------------------------------------------------------------------------------------------------------------|----------------------|----|-----|
|       |                                              |                                                                                                                      | I                    | II | III |
| 3a, b | <i>Peranema inflexum</i> Skuja               | Cell 32–25 µm long × 8–9 µm wide, colorless, various in shape; common                                                | +                    | –  | –   |
| 3c, d | <i>Euglena adhaerens</i> Matvienko           | Cell 80–85 µm long × 7.5 µm wide, cylindrical; chloroplasts discoid; not common                                      | +                    | +  | –   |
| 3e, f | <i>E. granulata</i> (G. A. Klebs) F. Schmitz | Cell 90–105 µm long × 25–30 µm wideW, spindle-shaped; flexible; chloroplasts disc-shaped with diplopyrenoids; common | +                    | –  | –   |
| 3g    | <i>E. cf. geniculata</i> Dujard.             | Cell 75–80 µm long × 17.5 µm wide, cylindrical; two stellate chloroplasts but slightly damaged; common               | +                    | –  | –   |
| 3h    | <i>Euglena</i> sp.                           | Cell 40–45 µm long × 25–30 µm wide                                                                                   | +                    | +  | –   |

Table 2. Continued.

| Figs  | Taxon                                                                                        | Taxonomical descriptions                                                                                                                                                                    | Sites and occurrence |    |     |
|-------|----------------------------------------------------------------------------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|----------------------|----|-----|
|       |                                                                                              |                                                                                                                                                                                             | I                    | II | III |
| 3i, j | <i>Euglenamorfis proxima</i> (P. A. Dang.)<br>M. S. Bennett & Triemer                        | Cell 52–55 µm long × 17.5–22.5 µm wide, spindle-shaped; chloroplasts discoid, lacking pyrenoids, several paramylon grains; common                                                           | +                    | +  | –   |
| 3k, l | <i>Euglenaria caudata</i> (E. F. W. Hübner) Karnkowska-Ishikawa,<br>E. W. Linton & Kwiat.    | Cell 100–110 µm long × 27.5–30.0 µm wide, spindle-shaped, chloroplasts discoid with diplopyrenoids; very common                                                                             | +                    | –  | –   |
| 3m    | <i>E. cf. clavata</i> (Skuja) Karnkowska-Ishikawa & E. W. Linton                             | Cell 35–40 µm long × 20.0–22.5 µm wide, club-shaped, chloroplasts discoid and irregularly lobed; common                                                                                     | +                    | +  | –   |
| 3n    | <i>Discoplastis spathirhyncha</i> (Skuja) Triemer                                            | Cell 65–70 µm long × 17.5–20.0 µm wide, variable in shape, usually from spindle to spinning top; chloroplasts numerous, discoid; not common                                                 | +                    | –  | –   |
| 3o, p | <i>Lepocinclis acus</i> (O. F. Müller) B. Marin & Melkonian                                  | Cell 75–150 µm long × 7.5–10.0 µm wide, spindle-shaped, narrow, elongated, rigid; chloroplasts numerous, discoid without pyrenoids; paramylon bodies numerous, long, rod-shaped; common     | +                    | +  | –   |
| 4a    | <i>L. fusiformis</i> (H. J. Carter) Lemmerm.                                                 | Cell 27.5–30 µm long × 18–20 µm wide, citroform, rigid; chloroplasts disc-shaped without pyrenoid, one large ring-like paramylon; common                                                    | +                    | –  | –   |
| 4b–d  | <i>L. oxyuris</i> (Schmarda) B. Marin & Melkonian                                            | Cell 132.5–140.0 µm long × 12.5–17.5 µm wide, cylindrical with hyaline caudus; chloroplasts small, numerous, discoid, two large paramylon body; common                                      | +                    | +  | –   |
| 4e, f | <i>L. texta</i> (Dujard.) Lemmerm.                                                           | Cell 37.5–40.0 µm long × 27.5–30.0 µm wide, ovoid, with small depression at apex; chloroplasts discoid without pyrenoid; paramylon bodies small, numerous, oval; common                     | +                    | +  | –   |
| 4g    | <i>Phacus anomalus</i> F. E. Fritsch & M. F. Rich                                            | Cell 25.0–27.5 µm long × 15–20 µm wide, irregular in shape, usually pear-shaped, with bent caudus; chloroplasts discoid, two paramylon bodies; not common                                   | +                    | –  | –   |
| 4h    | <i>Ph. crassus</i> Zakryś & M. Łukomska (delimited from the <i>Ph. longicauda</i> complex)   | Cell ca 60 µm long × 27.5–35.0 µm wide, widely oval, flattened with long straight cauda, with a short S-shaped fold; probably common                                                        | +                    | –  | –   |
| 4i, j | <i>Ph. cristatus</i> Zakryś & M. Łukomska (delimited from the <i>Ph. longicauda</i> complex) | Cell 50–60 µm long × 27.5–35.0 µm wide, widely oval, semi-flattened, with long fold, spirally twisted with long sharp caudus, chloroplasts discoid, small; paramylon body ring-like; common | +                    | –  | –   |
| 4k, l | <i>Ph. curvicauda</i> Svirenko                                                               | Cell 38.8–40.0 µm long × 25–30 µm wide, broadly ovoid, with longitudinal groove, short bent cauda; chloroplasts small, discoid, two large disc-like paramylon bodies; common                | +                    | +  | –   |
| 4m    | <i>Ph. orbicularis</i> K. Hübner                                                             | Cell 52.5–75.0 µm long × 30–50 wide, broadly ovoid, with sharp slightly bent cauda; chloroplasts small, discoid; one or two disc-like paramylon bodies; common                              | +                    | +  | –   |
| 4n    | <i>Ph. pleuronectes</i> (O. F. Müller) Nitzsch ex Dujard.                                    | Cell 75–80 µm long × 45–50 µm wide, broadly oval with short slightly bent caudus, slightly twisted; chloroplasts numerous, discoid, usually two disc-like paramylon bodies; common          | +                    | +  | –   |
| 4o    | <i>Trachelomonas cf. volvocina</i> (Ehrenb.) Ehrenb.                                         | Lorica 17.5–20.0 µm in diam., smooth, cell with two parietal chloroplasts, each with pyrenoids; common                                                                                      | –                    | –  | +   |



**Fig. 3.** a & b – *Peranema inflexum* Skuja, c & d – *Euglena adhaerens* Matvienko, e & f – *E. granulata* (G. A. Klebs) F. Schmitz, g – *E. cf. geniculata* Dujard., h – *Euglena* sp., i & j – *Eugleniformis proxima* (P. A. Dang.) M. S. Bennett & Triemer, k & l – *Euglenaria caudata* (E. F. W. Hübner) Karnkowska-Ishikawa, E. W. Linton & Kwiat., m – *E. cf. clavata* Karnkowska Ishikawa & E. W. Linton, n – *Discoplastis spathirhyncha* (Skuja) Triemer, o & p – *Lepocinclis acus* (O. F. Müller) B. Marin & Melkonian. Scale bars = 10  $\mu$ m.



**Fig. 4.** a – *Lepocinclis fusiformis* (H. J. Carter) Lemmerm., b–d – *L. oxyuris* (Schmarda) B. Marin & Melkonian, e & f – *L. texta* (Dujard.) Lemmerm., g – *Phacus anomalus* F. E. Fritsch & M. F. Rich, h – *Ph. crassus* Zakryś & M. Łukomska, i & j – *Ph. cristatus* Zakryś & M. Łukomska, k & l – *Ph. curvicauda* Svirenko, m – *Ph. orbicularis* K. Hübner, n – *Ph. pleuronectes* (O. F. Müller) Nitzsch ex Dujard., o – *Trachelomonas* cf. *volvocina* (Ehrenb.) Ehrenb. Scale bars = 10 μm.

(Saber, unpublished data) but Łukomska-Kowalczyk *et al.* (2015) recently delimited them as new species based on molecular and morphological. Here we identify them as new records for Egypt.

Detailed observations of the investigated species indicated that their chloroplast shape and structure were somewhat different from those

typically observed under laboratory conditions. The chemical properties of the habitats might be responsible for these changes, as suggested by results from Płachno *et al.* (2015), who found that the shape and arrangement of chloroplasts in some euglenophytes [particularly *Euglena viridis* (O. F. Müll.) Ehrenb. and *E. mutabilis* F. Schmitz]

sampled from heavily polluted streams in Poland varied markedly and did not always fit the original descriptions. Our present work, the first in-depth study of euglenoid species composition in Egypt, points to the need for further investigation of these isolated desert habitats.

## CONCLUSIONS

Twenty different euglenoids were identified from samples collected in the El-Farafra Oasis (The Western Desert). Four of these species (*Peranema inflexum*, *Euglena adhaerens*, *Phacus crassus*, *Ph. cristatus*) are new records for Egypt. All the species found occurred in agricultural and/or human-impacted habitats. More in-depth studies in Egypt will no doubt produce records of other interesting species, especially in isolated desert habitats.

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