#### **ORIGINAL ARTICLE**



# Tumor-Infiltrating Immunosuppressive Cells in Cancer-Cell Plasticity, Tumor Progression and Therapy Response

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#### Abstract

In most tumors, cancer cells show the ability to dynamically transit from a non-cancer stem-like cell to a cancer stem-like cell (CSC) state and vice versa. This cell plasticity has been associated with the epithelial-to-mesenchymal transition program (EMT) and can be regulated by tumor cell-intrinsic mechanisms and complex interactions with various tumor microenvironment (TME) components. These interactions favor the generation of a specific "CSC niche" that helps maintain the main properties, pheno-typic plasticity and metastatic potential of this subset of tumor cells. For this reason, TME has been recognized as an important promoter of tumor progression and therapy resistance. Tumors have evolved a network of immunosuppressive mechanisms that limits the cytotoxic T cell response to cancer cells. Some key players in this network are tumor-associated macrophages, myeloid-derived suppressor cells and regulatory T cells, which not only favor a pro-tumoral and immunosuppressive environment that supports tumor growth and immune evasion, but also negatively influences immunotherapy. Here, we review the relevance of cytokines and growth factors provided by immunosuppressive immune cells in regulating cancer-cell plasticity. We also discuss how cancer cells remodel their own niche to promote proliferation, stemness and EMT, and escape immune surveillance. A better understanding of CSC-TME crosstalk signaling will enable the development of effective targeted or immune therapies that block tumor growth and metastasis.

Keywords Immunosuppressive immune cells · Cancer-cell plasticity · Cancer stem cells · EMT · Metastasis · Therapy response

# Introduction

In recent years, various studies have been directed towards understanding cancer-cell plasticity, which is defined as the ability to switch between cancer stem-like cell (CSC) and non-CSC states [1], along with transdifferentiation capability [2, 3], and how these processes affect tumor growth and progression. In some tumors, the dynamic acquisition of CSC features has been associated with the induction of the epithelial-tomesenchymal transition (EMT) program, which promotes the motility and invasion of the epithelial-shaped cells and induces a mesenchymal-like phenotype that favors metastasis [4–6]. Although most studies have focused on tumor cellintrinsic mechanisms, such as genetic and epigenetic alterations, attention has recently turned towards understanding how tumor cell-extrinsic factors provided by the tumor microenvironment (TME) influence cancer progression and response to therapy [6-8].

The TME is composed of fibroblasts, mesenchymal stem cells, endothelial cells and pericytes, extracellular matrix (ECM) proteins, immune cells, including macrophages, myeloid-derived suppressor cells (MDSCs), natural killer (NK) cells, dendritic cells (DCs) and B and T cells, among others, and a variety of factors secreted by these cells [9]. It is well known that the communication between cells and their microenvironment is important for tissue homeostasis, the immune cells being responsible for host defense, tissue remodeling and the elimination of dying cells. Similarly, acquisition of mutations and the altered expression of some proteins in cancer cells activate a cytotoxic immune response that eradicates these malignant cells. However, there is growing evidence that TME can change in response to cancer cellderived signals to favor tumor growth and progression [7, 10]. In this scenario, tumor stromal cell-derived signaling contributes to sustain cancer cell proliferation, survival and

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invasion, as well as metastasis and angiogenesis [11]. Given that TME may display anti- or pro-tumoral properties, it has been suggested that the re-education of stroma cells towards anti-tumor activity may be an effective therapeutic strategy [12]. These bidirectional interactions favor the generation of a specific "CSC niche" that helps conserve the main properties of this subset of tumor cells [13] and preserves their phenotypic plasticity. As CSCs drive long-term tumor growth, are responsible for relapse after therapy [14], and are considered the most likely candidates for metastasis generation [15], a thorough understanding of CSC-TME crosstalk is fundamental for innovative therapeutic strategies development.

Tumors have evolved mechanisms to attenuate the effectiveness of T cells, whereby these cells become exhausted and dysfunctional, and therefore ineffective in attacking cancer cells [16, 17]. Tumors achieve this by creating an immunosuppressive network enriched in soluble mediators and specific immune cell populations. Components of TME, such as tumor-associated macrophages (TAMs), MDSCs and regulatory T (T<sub>reg</sub>) cells, which are highly effective in inhibiting T cell activation and proliferation, generate a tumor-promoting local environment that negatively influences immunotherapy [18, 19]. In addition, defects in antigen presentation, such as losing the expression of major histocompatibility complex (MHC) proteins in tumor cells, prevents them being recognized by cytotoxic T cells, precluding the elimination of these malignant cells [20]. Tumor cells and some immunosuppressive cells hijack the so-called 'immune checkpoint' pathways, which are important for maintaining self-tolerance and protecting tissues from damage. The best characterized immune-checkpoint receptors are cytotoxic T lymphocyteassociated antigen 4 (CTLA-4) and programmed cell death protein 1 (PD-1), which bind to CD80/CD86 and PD-L1/ PD-L2 ligands, respectively, to initiate checkpoint signaling and cytotoxic T cell inhibition. Tumor cells take advantage of these regulatory mechanisms and induce the expression of these ligands to prevent anti-tumor responses [21].

This review concerns itself with the relevance of cytokines and growth factors provided by immunosuppressive immune cells in regulating cancer-cell plasticity and tumor progression, and with how cancer cells remodel their own niche to promote their proliferation, stemness and EMT, and to escape immune surveillance. We also discuss recent therapeutic strategies that target the tumor-associated stroma to prevent disease progression.

# Immune Cells Contributing to the Tumor-Immunosuppressive Network

Several populations of tumor-immunosuppressive cells have been described in recent years. Here, we focus on the best characterized immune cells with an immunosuppressive function, notably TAMs, MDSCs and  $T_{reg}$  cells (Fig. 1).

#### **Tumor-Associated Macrophages (TAMs)**

Macrophages are terminally differentiated myeloid cells originating from monocytic precursors, whose functions are to eliminate infectious agents, promote wound healing and regulate adaptive immunity [22]. In mice, macrophages are characterized by the expression of markers such as CD11b, F4/80 and colony-stimulating factor-1 receptor (CSF-1R), and low levels of expression of Gr1, whereas, in humans, macrophages are identified by the expression of CD68, CD16, CD14, CD312, CD115, and other markers [23].

Macrophages have usually been classified into two subtypes according to their polarization state and their functional role. M1 or 'classically activated' macrophages are activated via T helper type I (Th1)-derived cytokines and/or bacterial products such as lipopolysaccharide (LPS). M1 macrophages secrete pro-inflammatory cytokines (IL-12, IL-1β, IL-6, IL-23 and tumor necrosis factor  $\alpha$  (TNF- $\alpha$ )), generate reactive oxygen species and nitric oxide (NO) and express a high level of MHC class II. These cells have a tumoricidal function. By contrast, M2 or 'alternatively activated' macrophages are activated in response to T helper type II (Th2)-derived cytokines, such as IL-4, IL-10, IL-13, and glucocorticoid hormones. These cells are characterized by a high level of expression of scavenging, mannose and galactose receptors, activation of the arginase (ARG1) pathway, production of IL-10, vascular endothelial growth factor (VEGF), matrix metalloproteinases (MMPs), and these cells facilitate tumor progression [24, 25]. Although the M1/M2 nomenclature has been useful for many years, it is now known that macrophages can change their polarity in response to environmental stimuli, acquiring intermediate plastic phenotypes. For this reason, it has been proposed that the TAM classification should be based on the anti-tumorigenic or pro-tumorigenic function of the macrophages [23].

The recruitment of monocytes into tumors is primarily regulated by tumor cell-derived cytokines, chemokines, and growth factors, such as CCL2, CCL7, CCL8, CCL3, CCL4, colony stimulating factor-1 (CSF-1), granulocyte-macrophage CSF (GM-CSF), macrophage-stimulating protein (MSP), platelet-derived growth factor (PDGF), VEGF-A, and transforming growth factor- $\beta$ 1 (TGF- $\beta$ 1). These cells may then differentiate into immunosuppressive M2-like macrophages in response to IL-4, IL-10, and IL-13 [10, 26].

In human colorectal, stomach and skin tumors, the presence of TAMs is associated with a favorable outcome, whereas in breast, prostate, ovarian, and cervical cancer, TAMs are linked to poor prognosis, suggesting that TAM infiltration may be a potentially prognostic marker of clinical outcomes. These disparate observations may be a consequence of the



Fig. 1 The tumor-immunosuppressive network limits cytotoxic immune-cell responses to cancer cells and promotes cancer-cell plasticity. This schema illustrates some of the mechanisms evolved by tumors to block cytotoxic immune cell responses and to promote tumor progression and metastasis. Immune cells, such as TAMs, MDSCs, and  $T_{reg}$  cells are recruited into the tumor and become educated by cancer (stem) cell-derived cytokines and chemokines to acquire pro-tumorigenic functions. These cancer and tumor-infiltrating immune cells disrupt immune surveillance through multiple mechanisms, including inhibition of antigen presentation by DCs, T cell proliferation, M1 macrophage polarization and NK-cell cytotoxicity. In addition, cancer cells and immuno suppressive TAMs inhibit cytotoxic T cell function by activating immune checkpoint pathways. Immunosuppressive cells enhance tumor progression by directly promoting EMT and CSC properties in cancer cells

through the secretion of cytokines such as TGF- $\beta$ , IL-6, EGF, among others, and support invasion and metastasis by secreting a variety of pro-tumorigenic cytokines and growth factors. Paracrine signal loops between CSCs and TAMs have been described, which enhance cancercell plasticity, immunosuppression and tumor progression. Hence, CSC-derived CSF-1 and GM-CSF promote TAM activation and, in turn, macrophage-derived EGF or CCL18 induce EMT, CSC state, and tumor cell invasion. Finally, immunosuppressive TAMs and MDSCs produce pro-angiogenic factors such as VEGF, TNF- $\alpha$ , ADM, PDGF, among others, to promote angiogenesis, or secrete MMPs, serine proteases, and cathepsins, leading to ECM alterations. MDSC, myeloid-derived suppressor cell; NK, natural killer; TAM, tumor-associated macrophage;  $T_{reg}$ , regulatory T cells

detection of macrophages with distinct differentiation features (pro-inflammatory M1-like vs. pro-tumoral M2-like cells). For instance, in colorectal cancer, pro-inflammatory TAMs attract T cells to the tumor site, promoting type-I T cell responses that lead to the inhibition of tumor-cell proliferation [27]. However, other studies demonstrated that TAMs support tumor growth and progression [23] by promoting angiogenesis [28], enhancing invasion and metastasis [29], and protecting tumor cells from apoptosis [30].

Immunosuppressive TAMs (M2-like) subvert anti-tumor immunity by eliminating M1-like immune responses and by impairing CD8<sup>+</sup> T cell activation through direct interaction with these cells or by secreting immunosuppressive cytokines such as IL-10, TGF- $\beta$ , ARG1, prostaglandins, and proteases [31] (Fig. 1). Macrophages express PD-L1/PD-L2 and CD80/CD86, which bind with the inhibitory receptors PD-1 and CTLA-4, respectively, resulting in the inhibition of T cell cytotoxic function. PD-L1 expression in TAMs is upregulated under hypoxic conditions, via hypoxia-inducible factor-1 $\alpha$ (HIF-1 $\alpha$ ) signaling [32, 33]. In addition, TAMs can express PD-1, which is associated with a reduction of their phagocytosis activity. PD-1<sup>+</sup> TAMs have a polarization M2-like and pro-tumorigenic function, and the blockade of PD-1/PD-L1 increases the phagocytosis activity of these cells, reducing tumor growth [34]. Therefore, anti-PD-1/PD-L1 therapy can block tumor growth by promoting CD8<sup>+</sup> T cell activation and anti-tumor activity of TAMs.

#### Myeloid-Derived Suppressor Cells (MDSCs)

MDSCs play an important role in tumor progression by evading the host's immune response. These cells arise as a consequence of the aberrant myelopoiesis associated with cancer, and are functionally defined as immunosuppressive immature myeloid cells [35]. Two main populations have been described: granulocytic or polymorphonuclear MDSCs (PMN-MDSCs), and monocytic MDSCs (M-MDSCs) [36]. PMN-MDSCs share phenotypic features with neutrophils, but are less phagocytic. By contrast, M-MDSCs exhibit a similar phenotype to that of inflammatory monocytes and differentiate into immunosuppressive macrophages and DCs under specific TME signals. In mice, PMN-MDSCs are identified by the expression of CD11b<sup>+</sup>Ly6G<sup>+</sup>Ly6C<sup>low</sup> and M-MDSCs as CD11b<sup>+</sup>Ly6G<sup>-</sup>Ly6C<sup>high</sup>, whereas, in humans, PMN-MDSCs are CD11b<sup>+</sup>CD14<sup>-</sup>CD15<sup>+</sup>CD33<sup>+</sup> cells, and M-MDSCs are CD11b<sup>+</sup>CD14<sup>+</sup> CD15<sup>-</sup>CD33<sup>+</sup>HLA<sup>-</sup>DR<sup>-/lo</sup> cells.

M-MDSCs are recruited to tumor sites by tumor cellderived cytokines, such as CCL2 and CCL5. In contrast, PMN-MDSCs are recruited by tumor cell-derived CXCL1, CXCL2, CXCL5, CXCL6, CXCL8, and CXCL12, as well as CCL2, CCL3, and CCL15 [37]. CXC chemokine receptor 2 (CXCR2) binds CXCL1, CXCL2, and CXCL5, among other chemokines, and plays an important role in the tumor recruitment of PMN-MDSCs. Several studies have shown that MDSCs disrupt immune surveillance mechanisms, such as T cell activation, DC antigen presentation, M1-like polarization, and NK cell cytotoxicity (Fig. 1). Factors implicated in MDSC suppressive activity include: i) the production of ARG1, which leads to the deprivation of arginine that is essential for several T cell functions; ii) inducible nitric oxide synthase (iNOS), which promotes NO synthesis; iii) production of reactive oxygen species (ROS); and iv) expression of cyclooxygenase-2 (COX-2), prostaglandin E2 (PGE2), IL-10, and TGF- $\beta$ , which induce severe anergy of effector immune cells. NO and ROS both induce T cell apoptosis and the nitration of cytokines and T cell receptor (TCR), which block T cell migration and cytotoxic activities against tumor cells. In addition, MDSCs upregulate the expression of PD-L1, blocking anti-tumor T cell-mediated activity via interaction with the PD-1 receptor of these cells [38, 39].

More recently, other MDSC functions have been described, such as the formation of a pre-metastatic niche [40], enhancement of tumor growth and invasion, and stimulation of angiogenesis [41] (see below). Furthermore, an elevated frequency of MDSCs is positively correlated with advanced disease and poor therapeutic response in patients with a range of cancers, including melanoma, colorectal, breast, bladder, thyroid and non-small-cell lung cancer (NSCLC) [42, 43]. Therefore, targeting MDSCs is emerging as an attractive approach to the design of new cancer treatments.

#### Regulatory T (T<sub>reg</sub>) Cells

 $T_{reg}$  cells are characterized by the expression of markers such as CD4, CD25 and forkhead box P3 (FOXP3) that plays an important role in  $T_{reg}$ -cell development and function [44]. Under physiological conditions,  $T_{reg}$  cells regulate the activation of T and B cells and maintain the homeostasis of cytotoxic lymphocytes [45]. In contrast,  $T_{reg}$  cells fulfill different functions during tumorigenesis, depending on the environmental stimuli, which means that they can be associated with poor prognosis, as in gastric, esophageal, pancreatic, liver, and breast carcinoma [46–50], or with improved survival, as in colorectal cancer [51].  $T_{reg}$ -cell recruitment in a range of human cancer types is mediated by tumor cell-derived CCL22 and CCL28, which attract CCR4- and CCR10-expressing  $T_{reg}$ cells, respectively [52–54] (Fig. 1).

Similar to MDSCs,  $T_{reg}$  cells facilitate tumor progression by interfering with the cytotoxic activity of T cells and suppressing tumor-antigen presentation [55]. In this regard,  $T_{reg}$ cells secrete immunosuppressive cytokines, such as IL-10, IL-35, and TGF- $\beta$ , and block the activity of T cells by direct interaction with these cells. In addition,  $T_{reg}$  cells express CD39 and CD73, which induce metabolic alterations that suppress cytotoxic T cells and/or NK activity [56–58].

# Impact of Tumor-Infiltrating Immunosuppressive Cells on Cancer-Cell Plasticity

Cancer-cell plasticity has been associated with the induction of the EMT program [4, 5], which promotes the acquisition of CSC properties. The induction of EMT is mediated by many growth factors and cytokines, such as TGF- $\beta$ , hepatocyte growth factor (HGF), epidermal growth factor (EGF), NF-KB, Notch, and Wnt [59]. Growing evidence indicates that TAMs, MDSCs and Treg cells enhance tumor progression by promoting EMT and CSCs properties in tumor cells (Fig. 1). Indeed, M2-like TAMs, through TLR4/IL-10 signaling pathway [60] or via TGF-B1 [61], promote EMT and CSC-like properties in pancreatic and hepatocellular carcinoma, respectively. TNF- $\alpha$ secreted by pro-inflammatory TAMs induces a switch from a differentiated to a dedifferentiated phenotype in melanoma cells, which escape immune surveillance and contribute to tumor relapse [62]. Furthermore, mesenchymal-like breast cancer cells induce immunosuppressive TAM activation via GM-CSF. In turn, these activated macrophages secrete CCL18, creating a positive feedback loop that promotes EMT and metastasis [63]. TAM-derived IL-6 activates the STAT3 pathway to promote

CSC self-renewal in breast and hepatocellular carcinoma [64]. Similarly, EGF released from TAMs activates the EGFR/STAT3 pathway of breast cancer cells, inducing the expression of SOX2 and other target genes involved in CSC state maintenance [65]. In addition, Lu and colleagues demonstrated that TAMs and tumor-associated monocytes physically interact with mouse mammary CSCs to support the maintenance of the stem-like state in this subset of tumor cells. EphA4 binding to its receptor on tumor cells resulted in the activation of Src and NF- $\kappa$ B signaling favoring the secretion of numerous cytokines involved in CSC maintenance [66].

In addition, CSCs can enhance the pro-tumor function of tumor-recruited macrophages. Hence, CSC-derived IFN $\beta$  induces the secretion of TAM-derived IFN-stimulated gene 15 (ISG15), which promotes CSC self-renewal and tumorinitiating features in pancreatic ductal adenocarcinomas (PDAC) [67]. Furthermore, PDAC CSCs secrete Nodal/ Activin A and TGF- $\beta$ 1, which induces an M2 phenotype that favors CSC self-renewal and invasion through the secretion of hCAP-18/LL-37 [68].

Several studies showed that MDSCs are also involved in the regulation of EMT and CSC features [69, 70]. MDSC-induced EMT occurs through the upregulation of COX-2 and the activation of the  $\beta$ -catenin/TCF4 pathway in nasopharyngeal cells, or through the stimulation of miR-101 expression in ovarian cancer cells [69, 71]. Furthermore, tumor-infiltrated MDSCs, which show signal transducer and activator of transcription 3 (STAT3) signaling activation, promote the stemness of pancreatic cancer cells by increasing the expression of SNAIL, SLUG, ZEB1, NANOG, and OCT-4 [72]. In this regard, studies based on a mouse model of breast cancer implicated MDSC-derived IL-6 in increasing tumor cell stemness [73]. Zhu et al showed that CXCR2<sup>+</sup> MDSCs are recruited and expanded during breast cancer progression, which promotes cancer-cell EMT via IL-6, and T cell exhaustion [74].

In ovarian cancer, the EMT-inducer transcription factor SNAIL induces CXCL1/2 expression through the NF-kB pathway and enhances MDSC tumor infiltration via CXCR2 [75]. CSCs stimulate the accumulation of MDSCs via the elevated production of G-CSF in mammary syngeneic tumor models. In turn, MDSCs trigger NOTCH signaling activation to enhance CSC properties in tumor cells, establishing a feedforward loop [76]. Accordingly, Peng and colleagues demonstrated that MDSCs endow breast cancer cells with stem-like features through IL6/STAT3 and NO/NOTCH crosstalk signaling, and their targeting may offer an opportunity to improve cancer therapies [77]. Although many studies have not distinguished between MDSC populations, a recent report demonstrated that M-MDSCs and PMN-MDSCs have opposite effects on tumor cells. Indeed, tumor-infiltrated M-MDSCs facilitate cancer-cell dissemination by inducing EMT/CSC properties. In contrast, pulmonary PMN-MDSCs support metastasis by reverting the EMT phenotype and promoting tumor-cell proliferation [78].

Finally, some reports have indicated that tumor-infiltrating  $T_{reg}$  cells contribute to cancer-cell plasticity. Indeed, the activation of AKT and MAPK signaling pathways by FOXP3<sup>+</sup>  $T_{reg}$ -derived IL-17, and by PGE2-mediated NF- $\kappa$ B activation, induce the expansion of the CSC population in colorectal cancer [79–81].

# **CSC/EMT** and Immune Evasion

A comprehensive study that immunophenotyped around 9000 samples of different solid cancers by their transcriptomic signature showed that tumor features may influence the characteristics of their immune infiltrate [82]. Tumors that are enriched for the EMT program, focal adhesion, ECM remodeling, angiogenesis, inflammation, and hypoxia genes, are associated with the infiltration of immunosuppressive cells, such as macrophages [82]. Several studies have revealed that CSC escape of immune surveillance occurs by several mechanisms. In this regard, CD44<sup>+</sup> CSCs of head and neck carcinomas express PD-L1, which binds to PD-1 receptor on T cells, blocking their cytotoxic function [83]. Dongre et al showed that epithelial-like breast tumors exhibit high levels of CD8<sup>+</sup> T cells and antitumor M1-like macrophages. In contrast, tumors arising from breast mesenchymal-like cells with an induced EMT, and a high level of expression of PD-L1, contain T<sub>reg</sub> cells, M2-like macrophages and exhausted CD8<sup>+</sup> T cells. These findings indicate that tumor cells with an induced EMT program promote the recruitment of immunosuppressive cells and escape immune surveillance [84]. Accordingly, ZEB1 expression relieves microRNA-200 (miR-200) repression of PD-L1 on NSCLC cells, leading to CD8<sup>+</sup> T cell immunosuppression and metastasis [85]. However, while epithelial tumor-bearing mice respond to anti-CTLA-4 immunotherapy, mesenchymal tumor-bearing mice are refractory to this treatment, suggesting that the immunosuppressive microenvironment of mesenchymal-like tumors makes them resistant to this therapy [84]. Accordingly, characterization of transcriptomic features in metastatic melanoma showed that resistant and non-responding melanomas to anti-PD-1 therapy display a gene signature characterized by the up-regulation of genes involved in EMT, angiogenesis and immunosuppression, characterized by monocytes and macrophages infiltration. These observations suggest that the mesenchymal and immunosuppressive phenotype is associated with innate anti-PD-1 resistance [86].

Another mechanism used by cancer cells to escape immune surveillance operates through alterations in the expression of MHC-I and MHC-II proteins. These proteins are required to elicit the immune responses by T lymphocytes, and MHC-I is downregulated in glioblastoma and breast CSCs [84, 87]. However, glioblastoma CSCs express various ligands of activating NK receptors, making them susceptible to NK-mediated cell cytotoxicity [88].

# Impact of Tumor-Infiltrating Immune Cells on Tumor Progression and Metastasis

Several studies have associated immunosuppressive TAMs with tumor-cell invasion and metastasis. The induction of angiogenesis is crucial to the development and growth of most solid tumors. TAMs produce a plethora of pro-angiogenic factors, including VEGF, TNF- $\alpha$ , adrenomedullin (ADM), PDGF, TGF- $\beta$ , and these cells are involved in promoting the formation of a vascular network during malignant progression in a mouse model of breast cancer [28]. In this regard, VEGF-A helps enhance vascularization and establishes an antiinflammatory microenvironment, which is characterized by the presence of M2-polarized macrophages in a model of skin carcinogenesis [89]. Consistent with this, TAM depletion in *Csf1*-null mice or by using liposome-encapsulated clodronate reduces angiogenesis in various tumor models [90].

In addition, immunosuppressive TAMs promote tumor-cell invasion and metastasis via secretion of MMPs, serine proteases, and cathepsins, which alter the ECM, modify cell-cell contacts and induce basal membrane disruption (Fig. 1). In this regard, cancer cell-secreted IL-4 induces a M2 macrophage polarization state characterized by increased cathepsin activity, and synergizes with IL-6 and IL-10 to promote tumor growth and invasion in pancreatic cancer [91, 92]. This synergy is dependent on the STAT3 and STAT6 interaction, which activates inositol-requiring enzyme  $1\alpha$  (IRE1 $\alpha$ ), leading to enhanced cathepsin secretion [92].

CSF-1 promotes the development of invasive and metastatic carcinomas by regulating the recruitment of TAMs at the tumor site [93]. CSF-1R inhibition abrogates TAM infiltration and enhances CD8<sup>+</sup> T cell recruitment, reducing cervical and mammary tumor growth and progression [94]. However, it was observed that glioma cell-supplied GM-CSF, IFNy, and CXCL10 promote TAM survival upon CSF-1R inhibition, although these TAMs lose M2 polarization and show enhanced phagocytosis activity [95]. Similarly, the inhibition of CCL2-CCR2 signaling specifically blocks macrophage recruitment within the tumor, which is associated with reduced metastasis and enhanced mouse survival [26]. A paracrine signaling loop between tumor-derived CSF-1 and macrophage-derived EGF has been linked to increased carcinoma cell invasion [96, 97]. Accordingly, M2-polarized TAMs by IL-4-expressing T<sub>H</sub>2-CD4<sup>+</sup> lymphocytes produced high levels of EGF, which enhance tumor-cell invasion, migration and lung metastasis by activating EGFR-signaling in breast cancer cells [98]. Therefore, these studies show that CD4<sup>+</sup> T cells can enhance breast cancer-cell dissemination and metastasis through their ability to regulate the pro-tumor properties of TAMs.

Several studies have described the impact of TAMs on the efficacy of chemotherapeutic agents (reviewed in [99]). TAMs protect myeloma cells from chemotherapy-induced apoptosis by inhibiting the activation of caspase-dependent apoptotic signaling [30]. These immune cells contribute to cisplatinchemoresistance of lung and colon CSCs through STAT3 and Hedgehog signaling by releasing milk-fat globulin epidermal growth factor-8 protein (MFG-E8) [100]. The juxtacrine activation of  $\alpha$ 4-integrin-expressing TAMs by vascular cell adhesion molecule-1 (VCAM-1)-positive breast cancer cells promotes tumor cell survival via the induction of PI3K/AKT signaling during lung metastasis development [101]. Interestingly, VCAM-1 also interacts with  $\alpha 4\beta 1$  in osteoclasts, contributing to bone metastasis [102]. Together, these findings demonstrate that tumor cells co-opt TAMs in order to survive therapies, and that the disruption of signaling axes between stromal and tumor cells may serve to prevent metastatic colonization.

On the other hand, several studies have also related MDSC function to tumor-cell migration, invasion and metastasis. The recruitment of MDSCs to the tumor site and to the premetastatic niche is induced by several cancer cell-derived chemokines, and is mediated by the expression CXCR2 in MDSCs, among others (see section 2.2). Hypoxia can enhance MDSC migration to the tumor site via HIF-1 $\alpha$ -mediated production of chemokines [103]. VEGF-A secreted by colorectal carcinoma cells stimulated TAMs to produce CXCL1, which favored the recruitment of circulatory CXCR2<sup>+</sup> MDSCs into pre-metastatic niche in order to promote liver metastasis [104]. Furthermore, hypoxia-induced lysyl oxidase (LOX), S100A8/A9, IL-6, and IL-10 are also implicated in the MDSC recruitment and the establishment of the pre-metastatic niche [105–107].

On the other hand, deletion of the gene encoding TGF- $\beta$ receptor II (Tgfbr2) in mammary carcinoma cells increases MDSC infiltration into tumors through SDF-1/CXCR4 and CXCL5/CXCR2 axes. These tumor-recruited MDSCs directly promoted tumor-cell invasion and metastasis through enhanced MMP and TGF- $\beta$  secretion [108]. Kumar and colleagues demonstrated that the upregulated expression of  $\Delta$ Np63 transcription factor is correlated with an increased number of MDSCs in triple-negative breast cancer (TNBC) patients. MDSC recruitment to the primary tumor and metastatic sites occurs through the direct  $\Delta Np63$ -dependent activation of the chemokines CXCL2 and CCL22. Likewise, CXCR2/CCR4 inhibitors reduce MDSC recruitment, angiogenesis and metastasis, and MDSCs secrete pro-metastatic factors, such as MMP9 and chitinase-3-like 1 to promote TNBC CSC function [109]. All these studies provide a rationale for the development of CXCR2 antagonists to prevent metastatic spread of tumor cells.

Finally, MDSCs are known to play a role in promoting angiogenesis. Hence, MDSC recruitment inhibition mediated by stem-cell factor (SCF) mRNA interference in breast tumor cells restores proliferation of tumor-infiltrating T cells, and the blockade of SCF receptor (cKit) prevents tumor-specific T cell anergy, T<sub>reg</sub> development, and angiogenesis [110]. In addition, MDSCs secrete Bombina variegata peptide 8 (Bv8), whose expression is upregulated by STAT3 signaling. STAT3 activation can also directly induce the secretion of VEGF and bFGF by MDSCs [111]. Blockade of Bv8 in combination with VEGF antibody inhibits angiogenesis and tumor growth [112]. Although VEGF antibody-mediated therapy has had some success in the clinic setting, tumors eventually become refractory to this treatment. MDSC recruitment could be a key mechanism mediating this resistance, as MDSCs can promote new vessel growth even in the presence of VEGF antibody [113, 114].

# Therapeutic Strategies for Targeting Tumor-Immune Microenvironment

Some therapeutic strategies have been directed towards targeting stromal components rather than tumor cells. Stromal cells have a relatively low mutation rate [13] and may be less susceptible to developing therapeutic resistance. In addition, taking advantage of the characteristic of the TME to display anti- or pro-tumoral properties, it has been suggested that their re-education may be an effective therapeutic strategy [115, 116]. As TAMs, MDSCs, and T<sub>reg</sub> cells play an important role in tumor progression and metastasis and their tumor infiltration is associated with poor prognosis in various tumor types, targeting these populations is proving to be an attractive therapeutic strategy [117–123] (Table 1).

Immune checkpoint inhibitors such as anti-PD-1, anti-PD-L1, and anti-CTLA-4 antibodies, which suppress the function of T cell-inhibitory receptors, have been developed as therapeutic strategies that increase the content of activated tumorspecific cytotoxic T cells [124] (Table 1). The first clinical trial with ipilimumab, an antibody that targets CTLA-4, showed longer overall survival to ~10 months in metastatic melanoma patients compared with patients not receiving ipilimumab therapy [125]. Additional clinical trials using CTLA-4 blocking drugs, either alone or in combination therapy are being performed on patients with advanced melanoma, NSCLC and breast cancer [126–128]. For instance, nivolumab, an anti-PD-1 receptor antibody, has been used alone or in combination with ipilimumab to treat patients with advanced melanoma, osteosarcoma, colorectal and renal carcinomas [129–133]. The 53% of melanoma patients had an objective response to combinatory therapy, all with tumor reduction of at least 80% [129] and longer overall survival compared with monotherapy [130]. Recently, FDA has approved a PD-1 blockade treatment for unresectable locally advanced and metastatic cutaneous SCCs with Cemiplimab [134]. However, most patients' clinical response was partial or short-lived, and a considerable percentage of patients suffered disease progression [135]. Checkpoint inhibitors that target PD-L1 have also been approved by FDA for the treatment of metastatic NSCLC, urothelial [136, 137], Merkel-cell carcinoma and advanced bladder cancer [138-140]. As some patients are intrinsically resistant or develop adaptive resistance after these treatments in several tumor types, inhibitors of additional immune checkpoint receptors, such as lymphocyte activation gene 3 (LAG3; CD223), and T cell immunoglobulin and mucin-containing molecule-3 (TIM3; CD366) [141] are currently being tested, alone or in combination with a variety of drugs, as new targets in cancer immunotherapy (Table 1).

Clinical trials with agonistic monoclonal antibody against CD40, a TNF receptor superfamily member that is expressed in DCs, B cells, monocytes and many nonimmune and tumor cells, have shown encouraging results in monotherapy and in combination with chemotherapy. The CD40 monoclonal antibody reverses immune suppression by activating APCs, promoting anti-tumor T cell responses and re-educating cytotoxic myeloid cells in lymphomas, melanomas, and pancreatic carcinomas [142] (Table 1).

On the other hand, chimeric antigen receptor-T cell (CAR-T) therapy is based on the ex vivo genetic modification of T cells in order to recognize tumor-associated antigens and kill target cells expressing these antigens [143]. CAR-T therapy has shown considerable success in treating leukemia but limited efficacy in solid tumors [143], except for neuroblastoma [144] and sarcoma [145]. These observations suggest that TME may block immune-driven tumor clearance by impairing CAR-T infiltration, activation or survival. Therefore, the removal of TME barriers to immune clearance may improve the efficiency of CAR-T-based therapies in solid tumors [146, 147].

Other TME-directed therapies are based on the neutralization of tumor-associated chronic inflammation [148]. The inhibition of the NF-KB pathway or immune-cell recruitment and/or function blockade through CSF-1R, CCR2 or CXCR2 inhibitors is being investigated in clinical trials [149]. For example, several CCR2 inhibitors (CCX872-B, PF-04136309, MLN1202, and BMS-813160) are currently undergoing clinical trials for the treatment of solid tumors (Table 1) [118]. In addition, CSF-1R inhibitors decrease tumor growth and increase survival in gliomas in preclinical trials, which are associated with macrophage reprogramming, but not with the depletion of these immune cells [95]. In contrast, treatment with CSF-1R inhibitor results in TAM depletion in preclinical models of breast cancer, without changes in the growth of primary tumors [149]. Moreover, PLX3397, a CSF-1R tyrosine kinase inhibitor, reduces macrophage

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Strategy	ıaıgdı	Agent	Biological function	Disease	Refs
Immune activation	CTLA-4	Ipilimumab	T-cell activation	Melanoma* Denolicitori triclet NICCT C. hencet concore	[125–128]
	PD-1	Nivolumab	T-cell activation	Freeninteat triatis. Nov.c.LC, 01 east cancer Metastatic melanoma*, NISCT C* sd PCC*	[129–133]
		Pembrolizumab		Mosceto and NCC*, Hodgkin lymphoma* Mensod and Advention automatics automatics and activity automatics automatics automatics automatics automatics	[124] [124]
	PD-L1	Atezolizumab	T-cell activation	Auvariced and inecasianty cutaneous SCC. Metastatic NSCLC* and UC*	[136, 137]
		Avelumab	Amplify anti-tumor immunity	Metastatic Merkel-cell* and UC*	[138]
	TIM3	Durvalumab Sym023 TSR-022	T-cell activation	Advanced bladder cancer* Phase I trials: advanced solid tumors and lymphomas	[139] [124]
		LY3321367 MBG453			
	LAG3	Sym022 TSB 02.2	T-cell activation	Phase I trials: advanced solid tumors and lymphomas	[124]
		15K-055 BMS-986016		Phase I trials: recurrent GBM and hematologic neoplasms	
Re-education	CD40	CD40 mAb	APCs and T-cell activation	Lymphoma, melanoma, pancreatic carcinoma	[142]
Macrophage-targeting	T cells CSF-IR CCR2	CAR-T PLX3397 CCX872-B MLN1202	re-cutudating synotry instrum cens Ex vivo genetic modification of T cells Macrophage infiltration reduction	Leukemia, large B cell lymphona, neuroblastoma, sarcoma Breast and prostate cancer, melanoma, GBM Phase I/II trials: PDAC, CRC and bone metastasis	[144–147] [118, 149–151] [118, 149]
	PI3K $\gamma$ in M2-like TAMs	BMS-813160 IPI-549 TG100–115	T-cell activation	HNSCC, PDAC, lung and breast cancer, melanoma	[118, 152]
	HRG HDAC	- TMP195 inhibitor	Macrophage polarization and angiogenesis Renolarizes TAMs Syneroizes with PD-1	Fibrosarcoma, pancreatic and breast cancer Breast cancer	[118, 155] [118, 156]
MDSCs-targeting	Class I HDAC STAT3	Entinostat AZD9150	Inhibition of MDSC activity	LLC and RCC Phase I trials: advanced HCC	[119–121] [119]
	CACRO	C87-X2	Blockade of MDSC recruitment	Phase II trials: pancreatic cancer, HNSCC, NSCLC and CRC	[110 122]
T <sub>reo</sub> -targeting	CD25	Daclizumab	Tree depletion	Breast cancer and melanoma	[123]
) )	CCR4	Mogamulizumab		Leukemia, lymphoma, lung and oesophageal cancer	[123]
	0.040	rr-04216000 MEDI6383	reduction of infinatio-suppressive activity		[(7]]
	GITR	MEDI1873		Advanced solid tumors	[123]
		TRX518 MK-1248			
	PI3Kδ	Parsaclisib	Increased CD8 <sup>+</sup> T-cell activity	Phase I trial: advanced solid tumors	[123]

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infiltration and enhances the efficacy of radiotherapy and immunotherapy [150, 151]. This drug is currently under clinical development for the treatment of glioblastoma and breast cancer patients. Given that different tumor types show distinct responses to CSF-1R inhibition, it has been suggested that TME-targeted therapies should be administered depending on tumor type and location in order to block tumor progression and metastasis development.

Strategies to promote the polarization switch of M2 macrophages to an anti-tumor M1 phenotype are also emerging as new therapeutic approaches (Table 1). In this regard, the inhibition of PI3K $\gamma$  in M2-like TAMs gives rise to T cell activation that suppresses growth of head and neck SCC (HNSCC), PDAC, lung and breast tumor [152]. Furthermore, activation of TLR3/Toll-IL-1 receptor domain-containing adaptor molecule 1 by Poly (I:C) accelerates M1-like macrophage polarization [153]. Polarization from an M2 to an M1 phenotype suppresses mammary tumor growth and angiogenesis [154], indicating that the re-education of TAMs could restore normal vasculature and block the pro-tumorigenic effects of TAMs. In accordance, histidine-rich glycoprotein (HRG) inhibits tumor growth and metastasis by inducing macrophage polarization and vessel normalization via downregulation of the placental growth factor (PIGF) [118, 155]. TMP195, a histone deacetylase (HDAC) inhibitor, repolarizes TAMs and synergizes with PD-1 to reduce tumor burden and metastasis in a breast cancer model [156].

Taken together, these studies suggest that a combination of therapies targeting immune evasion of tumor cells and those activating anti-tumor immune cell responses may provide optimal benefits for patients. However, these trials are still in the early stages and we do not fully understand how these drugs contribute to efficacy and overall survival. The long-term impact of these therapies on patient safety and survival also remains to be evaluated.

# **Concluding Remarks**

Studies in recent years have significantly advanced our understanding of the cytokines and molecular pathways regulating the crosstalk between TME components such as TAMs, MDSCs,  $T_{reg}$ , and cancer cells. These immunosuppressive cells and derived cytokines contribute to the generation of a tumor-immunosuppressive network that promotes tumor progression and negatively influences immunotherapy in a range of cancer types. Signals derived from these tumor-infiltrating immune cells help sustain cancer-cell proliferation, survival, and invasion, as well as metastasis and angiogenesis. In addition, cancer cells have evolved different mechanisms of immune evasion to avoid their recognition and/or attack by cytotoxic T cells or NK cells. These mechanisms include the recruitment of immunosuppressive cells that block the activity and proliferation of cytotoxic T cells, the downregulation of MHC-I protein expression, and/or the induction of PD-1 and CTLA-4 ligands, among others, for the activation of T cell-inhibitory immune checkpoint pathways.

Several studies have also shown that immune and cancercell crosstalk plays an important role in promoting the acquisition of CSC features, EMT, CSC self-renewal and maintenance. Likewise. CSC-derived factors stimulate the recruitment of these immunosuppressive cells into tumors. Given the important role of CSCs in driving long-term tumor growth, metastasis, and relapse after therapy, a thorough understanding of CSC-TME crosstalk is fundamental to the development of innovative therapeutic strategies. In this regard, therapies based on disrupting the pro-tumorigenic TME by blocking the recruitment of immunosuppressive TAMs and MDSCs to tumors have been developed. In addition, many studies have suggested that re-education of stromal cells, rather than their targeted ablation, may be an effective strategy for treating cancer. Future studies will allow us to establish combinatory therapies that block tumor growth and metastasis by targeting TME and cancer cells, and to increase the efficiency of currently immune checkpoint inhibitors.

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