

Development and Characterization of 35 SNP Markers in the Fat Greenling *Hexagrammos otakii*

Xianggang Gao^{1*}, Weidong Liu¹, Xiangbo Bao¹, Ying Xia²

¹Liaoning Key Laboratory of Marine Fishery Molecular Biology, Liaoning Ocean and Fisheries Science Institute, Dalian, China

²Dalian Eco-Environmental Monitoring Center of Liaoning Province, Dalian, China

Email: *xiangganggao@163.com, cnliu51@126.com, bobo_1325@126.com, xiaying517@163.com

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Abstract

Hexagrammos otakii is an important kind of economic fish species in East Asia. However, wild *H. otakii* resources have declined sharply in recent years as a result of human disturbance and habitat destruction. Thus, it is crucial to protect the current resources of *H. otakii*. In this study, 35 novel single nucleotide polymorphism (SNP) markers were developed based on restriction-site associated DNA sequencing. The results showed that the observed heterozygosity and expected heterozygosity ranged from 0.1875 to 0.6562 and 0.2679 to 0.5079, respectively. The minor allele frequency ranged from 0.1875 to 0.4375. Polymorphic information content ranged from 0.229 to 0.375. Six SNPs were found to be deviated significantly from the HWE ($P < 0.05$). These SNP markers will serve as a useful tool for genetic studies and population evaluation aimed at the conservation of *H. otakii*.

Keywords

Hexagrammos otakii, SNP, RAD, Genetic Diversity

1. Introduction

Hexagrammos otakii (Fat greenling), belonging to the family Hexagrammidae and genus *Hexagrammos*, is mainly distributed in the North Korea, Japan and China offshore [1]. *H. otakii* is an important kind of economic fish species in East Asia. *H. otakii* has high protein content (18.50%) and essential amino acid (7.25%), especially since the protein content is higher than most other fish [2]. Featuring meat quality, good taste, and high nutritional value, *H. otakii* is chal-

*Corresponding author.

lenged by continuously growing market requirements and reduction of fishery resources [3]. Thus, the demand for hypervariable molecular markers to provide a population-genetic perspective on conservation and management efforts of the spices becomes urgent [4] [5].

With the rapid development of next-generation sequencing technologies (NGS) [6], single nucleotide polymorphisms (SNPs) have been largely developed and widely used for genetic studies in aquaculture species such as *Sebastes schlegelii*, *Megalobrama terminalis* and *Coilia ectenes* [7] [8] [9]. For *H. otakii* resource conservation, we implemented restriction-site associated DNA (RAD) sequencing to facilitate the genetic evaluation which could provide a reference for the development of SNP markers.

2. Materials and Methods

2.1. Materials

In this study, a total of 32 *H. otakii* wild individuals were collected from northern Yellow Sea in China. Muscle tissues were sampled and stored in 95% molecular grade ethanol. Total genomic DNA was extracted from tissue samples using the TIANamp Marine Animals DNA Kit (Tiangen, Beijing, China) following the manufacturer's instructions.

2.2. Methods

All samples were used to construct the RAD libraries. Then, the libraries were sequenced on the Illumina HiSeq 4000 platform using 150 base pair (bp) paired-end reads. We trimmed the adapter sequences and low-quality reads (Phred score < 20) with Cutadapt [10]. Finally, 707,105 putative SNPs with the highest scores were generated, from which we randomly selected 100 candidate SNPs to test their applicability. The polymorphism of these candidate SNPs was further characterized in the samples mentioned above. Primer sequences for SNP loci were designed by Primer 5.0 software. The PCR reactions were conducted in 25 μ L volume containing 50 ng of genomic DNA, 1 \times PCR buffer, 1.5 mM MgCl₂, 0.2 mM dNTPs, 250 nM of each primer, and 1U of *Taq* polymerase (Takara, Dalian, China). The amplicons were checked by 1.0% agarose gel electrophoresis and sequenced on ABI 3730 DNA Analyzer (Applied Biosystems).

3. Results

The observed heterozygosity (H_o), expected heterozygosity (H_e), minor allele frequency (MAF), and P value representing the deviations from the Hardy-Weinberg equilibrium were estimated using POPGENE 32. The polymorphism information content (PIC) was calculated using Cervus 3.0 [11]. Among the test SNP markers in *H. otakii*, 35 polymorphic SNP markers were characterized in **Table 1**. The H_o and H_e were ranged from 0.1875 to 0.6562 and 0.2679 to 0.5079, respectively. The MAF ranged from 0.1875 to 0.4375. The PIC varied from 0.229 to 0.375, with an average of 0.3293. Six SNPs were found to be de-

viated significantly from the HWE ($P < 0.05$) (see **Table 2** for details). These results will be useful for understanding the genetic diversity of *H. otakii* to assist in the management of this germplasm resource.

Table 1. Summary information for the 35 SNP markers developed for the *H. otakii*.

Primer ID	Primer sequences	SNP type	SNP Position	H_o	H_e	MAF	PIC	P_{HWE}
SNP1	F: GTGCCGATGTTTATATCAGGCG R: CACATTTCTGTTTCGGGCTC	AT	97	0.3750	0.5060	0.3125	0.374	0.1368
SNP2	F: AATTCTGGTCCGGCAACAA R: AGCCCGAGATGAACAGAGAC	AG	118	0.3125	0.3095	0.3125	0.271	0.9550
SNP3	F: TGCAGGTGAACATAGATTTCCAG R: TGCTCCTCTGATGATGGCA	AG	104	0.2188	0.2892	0.2188	0.244	0.1510
SNP4	F: TCTTGTCCCTCCACTGTTGC R: TCCGGTCACAACCTCCACC	AG	95	0.2500	0.3472	0.2500	0.283	0.1022
SNP5	F: TCAAACAGGGAGAAACGGC R: TTACCACTTGAAGAGACTCCC	AG	73	0.5938	0.5074	0.1875	0.375	0.3283
SNP6	F: GGAACGTAGGGGAGTTGAGC R: CACGCACATACGCACATACG	TG	86	0.2500	0.2679	0.2500	0.229	0.6929
SNP7	F: TTCCCAGGTAAAGCCAAGC R: AATACCTCCTCGCTACTCG	AC	106	0.5312	0.4479	0.4063	0.349	0.2828
SNP8	F: AAGTGTTTCATCCGCTGGCTTA R: TCATTGTAGGTAGCCACACG	AC	90	0.6562	0.4836	0.2813	0.363	0.0400*
SNP9	F: GTTAAACTGAATATCAACAGGTG R: AACTTTGTAAAGTGCCTGG	TG	67	0.3750	0.3810	0.375	0.305	0.9277
SNP10	F: TGTTTGTGTTGCCACTATCAG R: GCTCAATGTCTTGAACCTGGG	AC	64	0.4688	0.5035	0.3125	0.373	0.6917
SNP11	F: TGTTAACGTTTCGGCCTGCTAG R: AGCAGGAAGTCAACCCACG	AC	76	0.4375	0.3472	0.4375	0.283	0.1291
SNP12	F: TTCTCTGTTACAGCAGCGC R: CAATGATAGCACAGGCAGC	AG	95	0.4688	0.5074	0.2500	0.374	0.6612
SNP13	F: TCTGACTGACATCGTGGATCAC R: ATGTTCTCAGGTTGCATCAGG	CT	70	0.2500	0.4365	0.2500	0.337	0.0136*
SNP14	F: CGTAGGCTGTTGCGTCTC R: TGTGCACAAGTATAGGCTGC	CT	78	0.2188	0.4241	0.2188	0.330	0.0051*
SNP15	F: ATTCAGCAGTGGACTTGGGG R: TTCACTCTTTCCCGGGCTTC	CT	118	0.1875	0.3095	0.1875	0.258	0.0206*
SNP16	F: TCTACTATGACGCTGTTACTCCAG R: ATGGTCAAGTCATCAGTGGC	AG	83	0.2812	0.3289	0.2813	0.271	0.3967
SNP17	F: ACACGATACTGCTTTGTACAC R: GCTTCTAGATTAGCTTAGCGC	CT	81	0.3125	0.3810	0.2813	0.305	0.2964
SNP18	F: ATGGATTTGCAGCACGGTTG R: AGCTAAAACCTGGCAGCTCA	AG	102	0.3750	0.5060	0.2500	0.374	0.1368
SNP19	F: CACTGGTGTGTTTGTGTCAAGG R: TCCAGCACCATCATCAGGTC	TG	67	0.3438	0.5074	0.3438	0.375	0.0637

Continued

SNP20	F: AAGAGAGAAAATACCAACACTG R: AAATGATCCTGGTCCTTGC	AG	70	0.5312	0.5035	0.2500	0.373	0.7511
SNP21	F: CGTTGGTTGTTAAGTGGCAG R: CAACTAAGGCAGAAGAGATAGACC	AG	64	0.5000	0.5079	0.2500	0.375	0.9284
SNP22	F: TTGGTTTTTGACGGTGTGCGC R: GGTGGAGTGGCTGTGATTCT	CT	99	0.4375	0.5079	0.2500	0.375	0.4255
SNP23	F: AGAGTGCTGAAGCCTCAACTG R: ACGTGATGGAAAGGGACACTG	TG	70	0.2812	0.3289	0.2813	0.271	0.3967
SNP24	F: TTCGGTTACGCGGGAGTTTGTGTC R: GTGGAACCCACAGGGTAACGTC	CT	117	0.4062	0.4836	0.4063	0.363	0.3570
SNP25	F: AGAGTCAAGACTAAGCGCG R: CTGTGTGAGTTTTTGGTGGC	CT	102	0.3438	0.4479	0.3438	0.344	0.1794
SNP26	F: CTGCCGTGACGAGTTCAGAT R: TGTGCGATCGTGTGACAGAA	CT	71	0.4688	0.4678	0.4063	0.354	0.9902
SNP27	F: CATAACGGACAAGAAAATGCCTC R: TTAACCAGTGACTGTCTGGAC	CT	74	0.5625	0.4762	0.3438	0.359	0.2965
SNP28	F: ACCGTGTGCTTTCAGTTCCTG R: GTAACACTGGAGGGTGGAGC	CT	100	0.5312	0.4678	0.3750	0.354	0.4340
SNP29	F: GCCACATTCTACACACTGTCCC R: TCAGCAGACGACAATGCAGC	CT	75	0.3438	0.4479	0.3438	0.344	0.1794
SNP30	F: TCCTTCTCTGGAGTCTTTGTGTC R: TGTGTTTTGAGGTTTTTGCAAGC	CT	74	0.3750	0.4107	0.3750	0.330	0.6145
SNP31	F: CTCTCCAAGCCTCAGCGAG R: ACTTCTCTCCTCCACTGCA	AG	113	0.5000	0.4901	0.3125	0.366	0.9073
SNP32	F: CAGGTCTCAAGCTCACCCCTGG R: ACTACTTGCGGATCACTTGTTGCC	TG	94	0.3125	0.3095	0.3125	0.258	0.9550
SNP33	F: AGCATTGCTTGATAATGACTGCC R: AGCGAATATCTGCGAAACGG	AT	98	0.5000	0.5079	0.2188	0.375	0.9284
SNP34	F: GCTGCCACAGTCACAGTTTCAC R: TCAGGTCCTCAGCATCAACGG	AG	72	0.2500	0.4583	0.2500	0.349	0.0087
SNP35	F: AGATGCAAGAGGGCCACTTCC R: AGGGATAGAGGCTGACGCCTG	CT	103	0.1875	0.3095	0.1875	0.258	0.0206

Table 2. The detailed information about the six SNPs found to be deviated significantly from the HWE ($P < 0.05$).

Primer ID	Genotype/observed value	Genotype/expected value	$\chi^2 (P_{HWE})$
SNP8	AA/9, AC/21, CC/2	AA/11.88, AC/15.23, CC/4.88	4.58 (0.0400)
SNP13	CC/18, CT/8, TT/6	CC/15.13, CT/13.75, TT/3.13	5.60 (0.0136)
SNP14	TT/19, C/T7, CC/6	TT/15.82, CT/13.36, CC/2.82	7.26 (0.0051)
SNP15	TT/23, CT/6, CC/3	TT/21.13, CT/9.75, CC/1.13	4.72 (0.0206)
SNP34	GG/17, GA/8, AA/7	GG/13.78, GA/14.44, AA/3.78	6.39 (0.0087)
SNP35	TT/23, CT/6, CC/3	TT/21.13, CT/9.75, CC/1.13	4.72 (0.0206)

4. Conclusion

Wild *H. otakii* resources have declined sharply in recent years as a result of human disturbance and habitat destruction. In this study, 35 novel SNP markers were developed based on restriction-site associated DNA sequencing. The results showed that the *Ho* and *He* ranged from 0.1875 to 0.6562 and 0.2679 to 0.5079, respectively. The minor allele frequency ranged from 0.1875 to 0.4375. Polymorphic information content ranged from 0.229 to 0.375. Six SNPs were found to be deviated significantly from the HWE ($P < 0.05$). These SNP markers will serve as a useful tool for genetic studies and population evaluation aimed at the conservation of *H. otakii*.

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Conflicts of Interest

The authors declare no conflicts of interest regarding the publication of this paper.

References

- [1] Ren, G., Liu, Q., Gao, T. and Yanagimoto, T. (2013) Population Demography and Genetic Structure of the Fat Greenling (*Hexagrammos otakii*) Inferred from mtDNA Control Region Sequence Analyses. *Biochemical Systematics and Ecology*, **47**, 156-163. <https://doi.org/10.1016/j.bse.2012.09.026>
- [2] Kang, B. and Wu, Y.F. (1999) Nutritional Composition Analyses of *Hexagrammos otakii*. *Marine Science*, **6**, 23-24.
- [3] Shen, Z., Wang, B., Hu, F., Liu, H., Guan, H. and Zheng, F. (2019) Isolation and Characterization of Microsatellite Loci in *Hexagrammos otakii* based on 2b-RAD Method. *Journal of Applied Ichthyology*, **36**, 55-61. <https://doi.org/10.1111/jai.13988>
- [4] F.A.O. (1993) Report of the Expert Consultation on Utilization and Conservation of Aquatic Genetic Resources. *FAO Fisheries Report*, No. 491, 58 p.
- [5] Khandaker, L., Akond, M., Liu, S., Kantartzi, S., Meksem, K., Bellaloui, N., Lightfoot, D. and Kassem, M. (2015) Mapping of QTL Associated with Seed Amino Acids Content in "MD96-5722" by "Spencer" RIL Population of Soybean Using SNP Markers. *Food and Nutrition Sciences*, **6**, 974-984. <https://doi:10.4236/fns.2015.611101>
- [6] Davey, J.W., Hohenlohe, P.A., Etter, P.D., Boone, J.Q., Catchen, J.M. and Blaxter, M.L. (2011) Genome-Wide Genetic Marker Discovery and Genotyping Using Next-Generation Sequencing. *Nature Reviews Genetics*, **12**, 499-510. <https://doi.org/10.1038/nrg3012>
- [7] Gao, X.G., Bao, X.B., Sun, W., Li, Y.F., Liu, Z.Y. and Liu, W.D. (2021) Isolation and Characterization of 38 SNP Markers for the Black Rockfish, *Sebastes schlegelii* by Next-Generation Sequencing. *Conservation Genetics Resources*, **13**, 413-416. <https://doi.org/10.1007/s12686-021-01226-3>

- [8] Yang, J.P., Li, X.H., Li, Y.F., Zhu, S.L., Chen, W.T. and Li, J. (2020) Isolation and Characterization of 30 SNP Markers in Guangdong Bream (*Megalobrama terminalis*) by Next-Generation Sequencing. *Conservation Genetics Resources*, **12**, 399-402. <https://doi.org/10.1007/s12686-020-01131-1>
- [9] Yu, A.Q., Shi, Y.H. and Yan, Y.L. (2019) Development and Characterization of 50 snp Markers in *Coilia ectenes*. *Conservation Genetics Resources*, **12**, 177-181. <https://doi.org/10.1007/s12686-019-01086-y>
- [10] Martin, M. (2011) Cutadapt Removes Adapter Sequences from High Throughput Sequencing Reads. *EMBnet Journal*, **17**, 10-12. <https://doi.org/10.14806/ej.17.1.200>
- [11] Kalinowski, S.T., Taper, M.L. and Marshall, T.C. (2007) Revising How the Computer Program CERVUS Accommodates Genotyping Error Increases Success in Paternity Assignment. *Molecular Ecology*, **16**, 1099-1106. <https://doi.org/10.1111/j.1365-294X.2007.03089.x>